Received: 13 January 2017 | Revised: 20 March 2017 | Accepted: 21 March 2017

DOI: 10.1111/gcb.13712

PRIMARY RESEARCH ARTICLE

WILEY Global Change Biology

Ecological regime shift drives declining growth rates of sea turtles throughout the West Atlantic

Karen A. Bjorndal[1] | Alan B. Bolten[1] | Milani Chaloupka[2] | Vincent S. Saba[3] | Cláudio Bellini[4] | Maria A. G. Marcovaldi[5] | Armando J. B. Santos[6] | Luis Felipe Wurdig Bortolon[6] | Anne B. Meylan[7,8] | Peter A. Meylan[8,9] | Jennifer Gray[10] | Robert Hardy[7] | Beth Brost[7] | Michael Bresette[11] | Jonathan C. Gorham[11] | Stephen Connett[12] | Barbara Van Sciver Crouchley[12] | Mike Dawson[13] | Deborah Hayes[13] | Carlos E. Diez[14] | Robert P. van Dam[15] | Sue Willis[16] | Mabel Nava[16] | Kristen M. Hart[17] | Michael S. Cherkiss[17] | Andrew G. Crowder[18] | Clayton Pollock[19] | Zandy Hillis-Starr[19] | Fernando A. Muñoz Tenería[20] | Roberto Herrera-Pavón[21] | Vanessa Labrada-Martagón[22] | Armando Lorences[23] | Ana Negrete-Philippe[24] | Margaret M. Lamont[25] | Allen M. Foley[26] | Rhonda Bailey[7] | Raymond R. Carthy[27] | Russell Scarpino[28] | Erin McMichael[28] | Jane A. Provancha[29] | Annabelle Brooks[30] | Adriana Jardim[5] | Milagros López-Mendilaharsu[5] | Daniel González-Paredes[31] | Andrés Estrades[31] | Alejandro Fallabrino[31] | Gustavo Martínez-Souza[31] | Gabriela M. Vélez-Rubio[31] | Ralf H. Boulon Jr[32] | Jaime A. Collazo[33] | Robert Wershoven[34] | Vicente Guzmán Hernández[35] | Thomas B. Stringell[36] | Amdeep Sanghera[37] | Peter B. Richardson[37] | Annette C. Broderick[36] | Quinton Phillips[38] | Marta Calosso[39] | John A. B. Claydon[38] | Tasha L. Metz[40] | Amanda L. Gordon[41] | Andre M. Landry Jr[40] | Donna J. Shaver[42] | Janice Blumenthal[43] | Lucy Collyer[43] | Brendan J. Godley[36] | Andrew McGowan[36] | Matthew J. Witt[44] | Cathi L. Campbell[1] | Cynthia J. Lagueux[1] | Thomas L. Bethel[45] | Lory Kenyon[46]

[1]Archie Carr Center for Sea Turtle Research and Department of Biology, University of Florida, Gainesville, FL, USA

[2]Ecological Modelling Services Pty Ltd, University of Queensland, St Lucia, QLD, Australia

[3]NOAA National Marine Fisheries Service, Northeast Fisheries Science Center, Geophysical Fluid Dynamics Laboratory, Princeton, NJ, USA

[4]Centro TAMAR-ICMBio, CLBI – Parnamirim, Rio Grande do Norte, Brazil

[5]Fundação Pró TAMAR, Salvador, Bahia, Brazil

[6]Fundação Pró-TAMAR, Pernambuco, Brazil

[7]Florida Fish and Wildlife Conservation Commission, Fish and Wildlife Research Institute, St. Petersburg, FL, USA

[8]Smithsonian Tropical Research Institute, Washington, DC, USA

[9]Natural Sciences Collegium, Eckerd College, St. Petersburg, FL, USA

[10]Bermuda Turtle Project, Flatts, Bermuda

 © 2017 John Wiley & Sons Ltd

[11]Inwater Research Group, Jensen Beach, FL, USA
[12]Family Island Research and Education, Newport, RI, USA
[13]Geronimo Program, St. George's School, Newport, RI, USA
[14]DRNA-PR, San Juan, Puerto Rico
[15]Chelonia Inc, San Juan, Puerto Rico
[16]Sea Turtle Conservation Bonaire, Kralendijk, Bonaire, Dutch Caribbean
[17]U.S. Geological Survey, Wetland and Aquatic Research Center, Davie, FL, USA
[18]Cherokee Nation Technologies, NSU Center for Collaborative Research, Davie, FL, USA
[19]National Park Service, Christiansted, St. Croix, Virgin Islands
[20]Facultad de Agronomía y Veterinaria, Universidad Autónoma de San Luis Potosí, San Luis Potosí, México
[21]El Colegio de la Frontera Sur -Unidad Chetumal, Chetumal, Quintana Roo, México
[22]Facultad de Ciencias, Universidad Autónoma de San Luis Potosí, San Luis Potosí, México
[23]Dirección de Ecología Municipio de Solidaridad, Quintana Roo, México
[24]Parque Xcaret, Municipio de Solidaridad Playa del Carmen, Quintana Roo, México
[25]US Geological Survey, Wetland and Aquatic Research Center, Gainesville, FL, USA
[26]Florida Fish and Wildlife Conservation Commission, Fish and Wildlife Research Institute, Jacksonville Field Laboratory, Jacksonville, FL, USA
[27]US Geological Survey, Florida Cooperative Fish and Wildlife Research Unit, Gainesville, FL, USA
[28]Florida Cooperative Fish and Wildlife Research Unit, Department of Wildlife Ecology and Conservation, University of Florida, Gainesville, FL, USA
[29]Environmental Services, Integrated Mission Support Services, Kennedy Space Center, Florida, USA
[30]Cape Eleuthera Institute, Rock Sound, Eleuthera, The Bahamas
[31]Karumbé, Montevideo, Uruguay
[32]National Park Service, St. John, Virgin Islands
[33]U.S. Geological Survey, North Carolina Cooperative Fish and Wildlife Research Unit, Department of Applied Ecology, North Carolina State University, Raleigh, NC, USA
[34]Boca Raton, FL, USA
[35]APFFLT-CONANP, Campeche, México
[36]Centre for Ecology and Conservation, University of Exeter, Penryn, Cornwall, UK
[37]Marine Conservation Society, Ross-on-Wye, Herefordshire, UK
[38]Department of Environment and Coastal Resources, National Environment Centre, Providenciales, Turks and Caicos Islands
[39]The School for Field Studies, Center for Marine Resource Studies, South Caicos, Turks and Caicos Islands
[40]Texas A&M University at Galveston, Galveston, TX, USA
[41]Environmental Institute of Houston, University of Houston – Clear Lake, Houston, TX, USA
[42]Padre Island National Seashore, Corpus Christi, TX, USA
[43]Department of Environment, Grand Cayman, Cayman Islands
[44]Environment and Sustainability Institute, University of Exeter, Penryn, Cornwall, UK
[45]Hope Town, Abaco, The Bahamas
[46]Elbow Reef Lighthouse Society, Abaco, The Bahamas

Correspondence
Karen A. Bjorndal, Archie Carr Center for Sea Turtle Research and Department of Biology, University of Florida, Gainesville, FL, USA.
Email: bjorndal@ufl.edu

Abstract

Somatic growth is an integrated, individual-based response to environmental conditions, especially in ectotherms. Growth dynamics of large, mobile animals are particularly useful as bio-indicators of environmental change at regional scales. We assembled growth rate data from throughout the West Atlantic for green turtles, *Chelonia mydas*, which are long-lived, highly migratory, primarily herbivorous mega-consumers that may migrate over hundreds to thousands of kilometers. Our dataset, the largest ever compiled for sea turtles, has 9690 growth increments from 30 sites from Bermuda to Uruguay from 1973 to 2015. Using generalized additive mixed models, we evaluated covariates that could affect growth rates; body size, diet, and year have significant effects on growth. Growth increases in early years until 1999, then declines by 26% to 2015. The temporal (year) effect is of particular interest

because two carnivorous species of sea turtles—hawksbills, *Eretmochelys imbricata*, and loggerheads, *Caretta caretta*—exhibited similar significant declines in growth rates starting in 1997 in the West Atlantic, based on previous studies. These synchronous declines in productivity among three sea turtle species across a trophic spectrum provide strong evidence that an ecological regime shift (ERS) in the Atlantic is driving growth dynamics. The ERS resulted from a synergy of the 1997/1998 El Niño Southern Oscillation (ENSO)—the strongest on record—combined with an unprecedented warming rate over the last two to three decades. Further support is provided by the strong correlations between annualized mean growth rates of green turtles and both sea surface temperatures (SST) in the West Atlantic for years of declining growth rates ($r = -.94$) and the Multivariate ENSO Index (MEI) for all years ($r = .74$). Granger-causality analysis also supports the latter finding. We discuss multiple stressors that could reinforce and prolong the effect of the ERS. This study demonstrates the importance of region-wide collaborations.

KEYWORDS
Caretta caretta, *Chelonia mydas*, ecological regime shifts, *Eretmochelys imbricata*, multivariate ENSO index, sea surface temperature, seagrass, somatic growth rates

1 | INTRODUCTION

Marine systems have undergone great changes in ecosystem function and species distribution and abundance in the Anthropocene (Alheit, 2009; Halpern et al., 2015; Jackson et al., 2001; Lotze et al., 2006; McCauley et al., 2015). Some of these changes have resulted in or resulted from ecological regime shifts (ERS), defined by Conversi et al. (2015) as "dramatic, abrupt changes in the community structure that are persistent in time, encompass multiple variables, and include key structural species—independently of the mechanisms causing them." Many studies have evaluated the changes that result from ERS in structure and function of ecosystems; biogeography, phenology, and abundance of species; and foodwebs or trophodynamics (references in Alheit & Bakun, 2010; Rocha, Peterson, & Biggs, 2015; Young et al., 2015). However, fewer studies have addressed long-term physiological changes at the individual level across regional landscapes in this era of changing seas. Here, we evaluate somatic growth dynamics of the green turtle (*Chelonia mydas*) throughout the West Atlantic across more than four decades (1973–2015).

Somatic growth rates of ectotherms are valuable bio-indicators of environmental change because their growth dynamics are strongly influenced by environmental conditions and are an integrated response to changes in these conditions. Sea turtles are long-lived, highly migratory mega-consumers and are therefore excellent models for such environmental monitoring. Green turtles, hawksbills (*Eretmochelys imbricata*), and loggerheads (*Caretta caretta*) spend decades in neritic habitats growing to sexual maturity. During this immature period, individuals may move hundreds to thousands of kilometers among foraging grounds (Musick & Limpus, 1997).

Many of the authors of this study collaborated on earlier studies of somatic growth dynamics in West Atlantic hawksbills (Bjorndal et al., 2016) and Northwest Atlantic loggerheads (Bjorndal et al., 2013) based on capture–mark–recapture data and using a similar modeling approach. These studies revealed that growth rates for hawksbills and loggerheads exhibited similar, continuing declines beginning in 1997. The same pattern of decline was reported for North Atlantic loggerheads based on a different technique (skeletochronology) and a different sample of loggerheads (Avens et al., 2015). Hawksbills and loggerheads are primarily carnivorous, although they feed on different types of prey. Hawksbills feed mostly on sponges, corallimorpharians, zoanthids, and sea anemones associated with coral reefs (references in Krueger, Chaloupka, Leighton, Dunn, & Horrocks, 2011). Loggerheads prey most commonly on slow-moving or sessile, hard-shelled benthic invertebrates (Hopkins-Murphy, Owens, & Murphy, 2003).

In 1997/1998, an ERS occurred in the Atlantic as a result of a synergy between the abrupt warming from the strongest El Niño Southern Oscillation (ENSO) event ever recorded and the unprecedented warming rate over the last two to three decades (Beaugrand et al., 2015; IPCC, 2014; Martinson, Stammerjohn, Iannuzzi, Smith, & Vernet, 2008; Reid & Beaugrand, 2012; Wijffels, Roemmich, Monselesan, Church, & Gilson, 2016). The decline in hawksbill and loggerhead growth rates may have been a response to this ERS. A study of somatic growth dynamics of the primarily herbivorous green turtle would reveal the extent to which patterns of regional changes in productivity hold across trophic levels. If growth in green turtles follows the same pattern, the probability that the growth dynamics of all three species are responses to widespread climatic drivers and an ERS would be greatly increased. Therefore, we assembled growth rate data for West Atlantic green turtles resulting in the largest ($n = 9{,}690$ growth increments, longest (from 1973 through 2015), and most widespread (from Bermuda to Uruguay) dataset ever compiled for sea turtles.

In this study, we have three objectives: (1) evaluate West Atlantic green turtle growth dynamics with generalized additive mixed models, (2) compare the temporal dynamics of green turtles with those of West Atlantic hawksbills and North Atlantic loggerheads, and (3) explore relationships of temporal growth trajectories with Multivariate El Niño Southern Oscillation Index (MEI) and sea surface temperature (SST). These drivers were selected because they are the most likely drivers of the ERS in the late 1990s (Beaugrand et al., 2015; Martinson et al., 2008; Reid & Beaugrand, 2012).

2 | MATERIALS AND METHODS

2.1 | Data assembly

Green turtle growth rate data were combined from 30 projects in the West Atlantic (Figure 1). Some of these data have been published in studies for individual sites, but never in regional assessments. Turtles were captured by a variety of methods in foraging areas in neritic habitats and not on nesting beaches. Turtles were tagged, usually with flipper tags, for individual identification. Data used in this study are capture date and location (latitude/longitude), carapace length (CL, the most common measure of body size in sea turtles), and primary diet at each site. Sex is known for a small fraction of individuals so is not used in our analyses. Body size for each growth increment is the average of CL at capture and recapture (Chaloupka & Limpus, 1997). Negative growth rates, which result from either measurement error or damage to carapace margins, are included in analyses to avoid systematic bias.

When the growth data were first assembled, durations (time-at-large) of the growth increments varied from 1 to 7,636 days. Including growth increments with short or long durations can introduce substantial error. Short durations may only capture the fastest or slowest of seasonal growth rates, resulting in large errors when extrapolated to estimates of annual growth, or the change in size may be so small that measurement error is a large proportion of actual growth. During long durations, average CL may not represent a good estimate of body size for the interval. To set the minimum and maximum durations for our analyses, we followed Bjorndal et al. (2016) to determine the limits within which duration did not significantly affect our growth model. We created a dataset in which 60 days was the minimum duration (n = 9,690) and, based on the generalized additive mixed model (below), determined that 330 and 1,644 days were the minimum and maximum values. Our minimum value is the same as the standard that has been used for many years in sea turtle studies (Chaloupka & Limpus, 1997), giving further support to the standard minimum. To increase sample size, successive growth increments for individual turtles below the 330 days limit were combined to exceed the minimum duration when possible.

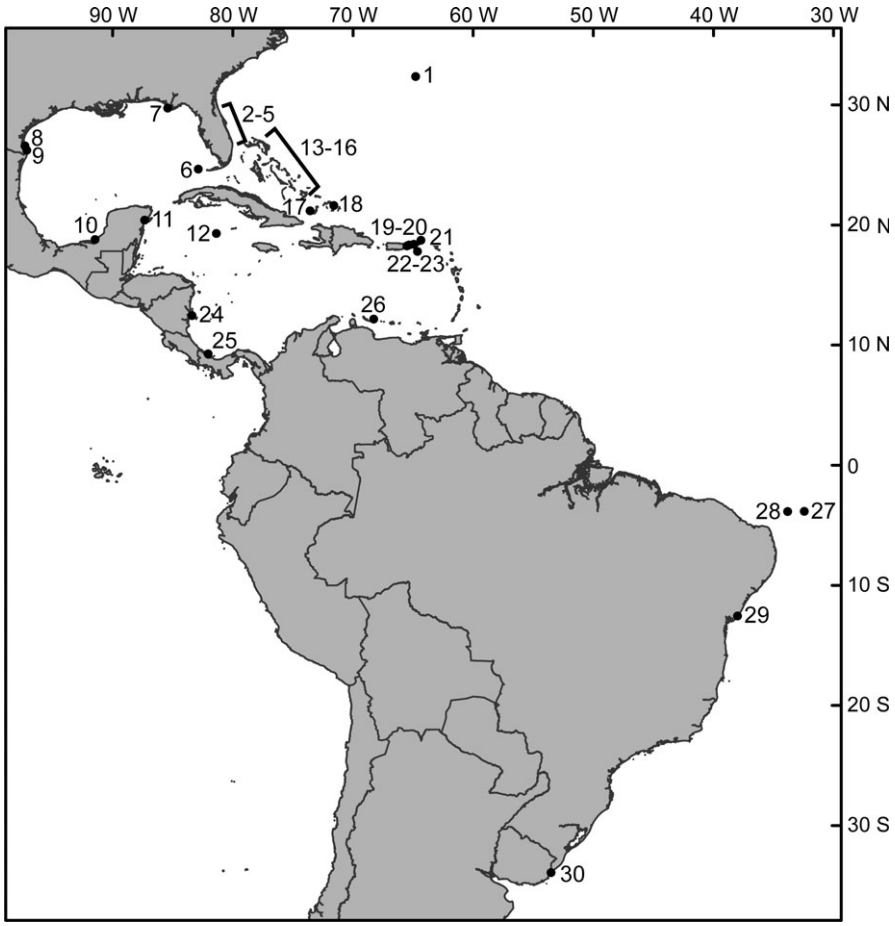

FIGURE 1 Location of study sites based on dataset with recapture durations ≥330 days and ≤1644 days (n = 6,201). 1 = Bermuda (n = 845); 2–5 = Florida East Coast, USA (n = 878); 6 = Dry Tortugas, Florida, USA (n = 53); 7 = St. Joseph Bay, Florida, USA (n = 64); 8 = Mansfield Channel, Texas, USA (n = 14); 9 = Laguna Madre, Texas, USA (n = 15); 10 = Campeche, México (n = 17); 11 = Akumal, México (n = 80); 12 = Cayman Islands (n = 9); 13–16 = Bahamas North & Central (n = 1,111); 17 = Great Inagua, Bahamas (n = 1,119); 18 = Turks and Caicos Islands (n = 15); 19–20 = Puerto Rico (n = 284); 21 = British Virgin Islands (n = 7); 22–23 = US Virgin Islands (n = 95); 24 = Pearl Cays, Nicaragua (n = 7); 25 = Panama (n = 36); 26 = Bonaire (n = 191); 27 = Fernando de Noronha, Brazil (n = 1,206); 28 = Atol das Rocas, Brazil (n = 89); 29 = Praia do Forte, Brazil (n = 39); 30 = Uruguay (n = 27)

2.2 | Statistical methods

Generalized additive nonparametric regression models with fixed and random effects—often referred to as generalized additive mixed models (GAMM)—were used to explore somatic growth rates. This modeling approach allows for flexible specification of both error and link functions, enables arbitrary specification of the functional form for each continuous covariate included in the model, and accounts for mixed effects from multiple measurements on the same sampling unit such as location (Fahrmeir & Lang, 2001). Our model used scaled Student-t (scat) likelihood based on findings from a gamboostLSS model as in Gilman, Chaloupka, Peschon, and Ellgen (2016) that showed Student-t likelihood is better than Gaussian for our model.

The GAMMs were fitted using the following: (1) thin plate regression splines to model nonlinear covariate effects, (2) a two-dimensional Duchon-spline surface smoother to account for structured spatial effects attributable to the geospatial location (latitude, longitude) of each project site, (3) a tensor product of a 2D Duchon-spline surface and a time effect with cubic regression spline basis to account for any spatial trend in time (Marra, Miller, & Zanin, 2012), where time is blocks of years (=epochs), and (4) project-specific heterogeneity incorporated as a random effect term to account for the multilevel sampling structure of the dataset. This spatially explicit GAMM is generally referred to as a geoadditive GAMM (Kammann & Wand, 2003). All GAMM models were fitted using the MGCV package for R (Wood & Scheipl, 2014) with the smoothness parameters estimated using REML (Wood, 2006).

We use a mixed longitudinal sampling design (sampling with partial replacement); 1318 (33%) of 3958 individual turtles were recaptured more than once. In our GAMM analyses, we assess six fixed effects and one random effect (project collecting the data, $n = 30$) on one response variable (somatic growth rate). Of the six fixed effects, two (diet and CL type) are each four-level factors. Diet is the primary diet for the site: seagrass, algae, seagrass/algae mix, and omnivorous. CL type is the specific CL metric used (see Appendix S1). The other four fixed effects are continuous covariates (mean CL of growth increment, mean year of growth increment, duration of growth increment, and location on a latitude/longitude surface or a location/temporal interaction term). Mean CL is the arithmetic mean of straight CL notch to tip (SCLnt, see Fig. S1-1 in Appendix S1) at initial capture and recapture. Mean year is the calendar year of the midpoint of the recapture interval. This approach introduces little error in calendar year assignment because 72% of growth records had durations <2 years. Recapture interval was included to evaluate any bias from variable durations. For the spatiotemporal interaction, we use an interaction term of location by epoch. The four epochs have nearly equal sample sizes based on mean year (1974–1999, 2000–2006, 2007–2010, 2011–2015). Number of growth increments in each epoch is 1470, 1421, 1486, and 1824, respectively. We conducted two GAMM analyses—a spatial model and a spatiotemporal model—to explore the importance of spatiotemporal interaction. In GAMM analyses, each covariate is conditioned on all other covariates. For example, any differences in CL of turtles in different regions or different years would be accounted for in assessments of spatial or temporal effects.

The R code for the spatiotemporal model is as follows: mgcv(data. gam <- gam(grow.rate ~ diet + cl.type + s(mean.size) + s(mean.year, $k = 4$) + s(duration) + te(lon, lat, by=decade, bs="ds," m=c(1,.5)) + s (project, bs="re"), family=scat(link="identity"), method="REML")).

Annualized mean growth rates are expressed as standardized values ([Annual value − mean of annual values]/*SD* of annual values) to allow direct comparison among the three sea turtle species. To evaluate whether the significant region-wide effect of mean year on growth rates was related to the two drivers proposed for the Atlantic ERS beginning in the mid-1990s (ENSO and ocean heating), we related annualized mean growth rates generated from our GAMM analysis to the MEI and SST. We selected the MEI to represent ENSO because it is currently considered the most representative index (Mazzarella, Giuliacci, & Scafetta, 2013) and reflects ecological changes well because it integrates six variables in the eastern tropical Pacific Ocean as a proxy for the ENSO: SST, surface air temperature, sea-level pressure, two components of surface winds, and total cloudiness of the sky (Mazzarella et al., 2013). Climate teleconnection between the Pacific and Atlantic is strong with the ENSO affecting Atlantic SST, rainfall, and associated regional-scale ocean-atmosphere anomalies in our study region (Giannini, Chiang, Cane, Kushnir, & Seager, 2001; Gouirand, Moron, Hu, & Jha, 2014; Spillman, Alves, & Hudson, 2011) and Atlantic warming possibly triggering ENSO events in the Pacific (Ham, Kug, Park, & Jin, 2013).

We sourced MEI bimonthly data from 1950 to present (http://www.esrl.noaa.gov/psd/enso/mei/table.html) and annualized the bimonthly index to an annualized index. We then ran a GAMM with autoregressive (AR1) error to reveal any underlying annual trend since 1950 and lag plotted the GAMM trend MEI against annualized mean growth rates for 0- to 11-year lags with *astsa* package for R (Stoffer, 2014). We followed a similar approach with SST data. Because of the variation in temporal and spatial coverage of our growth data, rather than using the SST values for the entire region, we used mean SST values from 625 km^2 around three sites (Bermuda; Inagua, The Bahamas; and Fernando de Noronha, Brazil; Figure 1). These three sites represent 53% of the growth increments in our study, 100% of the temporal range, and the latitudinal range of 93% of our study sites. SST data were sourced from NOAA OISST (Optimum Interpolation Sea Surface Temperature) AVHRR (Advanced Very High Resolution Radiometer) satellite data (25-km × 25-km (1/4 degree) https://www.ncdc.noaa.gov/oisst). SST data begin in 1982 when the satellite started recording data.

We also explored the relationship between somatic growth rates and MEI using a statistical forecasting approach. For instance, does an environmental driver such as MEI improve the forecasting performance of expected somatic growth rates for West Atlantic green turtles? A common test of this forecasting performance is Granger-causality analysis (Enders, 1995; Triacca, 2005). This analysis in no way assesses true causality and refers only to forecast ability while also assuming a linear dependence between the response variable

and the predictor (Mariusz, 2015). See Appendix S1 for details of the Granger-causality analysis.

3 | RESULTS

3.1 | Dataset and GAMM results

Green turtles were sampled in mixed stock foraging aggregations that include turtles from all five Atlantic regional management units (Bjorndal & Bolten, 2008; Wallace et al., 2010). Our initial dataset (n = 9,690) with a 60-days minimum duration for growth increments was revised to a dataset with durations from 330 to 1,640 days. Our final dataset has 6201 growth increments for 3958 individual green turtles. Number of growth increments for individual turtles varies from 1 to 10 with a mean ± *SD* of 1.6 ± 1.1. Growth rates from all growth increments vary from −0.9 to 11.9 cm/year with a mean ± SD of 3.4 ± 2.0 cm/year. Green turtles were captured from 1973 through 2015, and mean year of growth increments is from 1974 through 2015. SCLnt values from all turtle captures (n = 12,402) vary from 23.2 to 117.0 cm, and mean SCLnt values for all growth increments (n = 6,201) range from 24.6 to 117.0 cm. Mean CL of growth increments did not change over time (linear regression, n = 6,201, p = .289).

In a comparison of the spatial-only and the spatiotemporal GAMM analyses, the spatiotemporal interaction is significant for each of the four epochs (p < .007), so we only present results from the spatiotemporal model (Figure 2, Fig. S2-1 in Appendix S2). The spatiotemporal model explains 34.3% of the model deviance and is an adequate fit to the data with significant nonlinear effects. Including the spatiotemporal interaction in the model only increases the deviance explained by 1.3%. Of the seven covariates in the model, four of the six fixed effects and the one random effect (projects) are significant.

Mean CL is a significant fixed effect (p < .0001; Figure 2a). Growth rates initially increase from 25 to ~40 cm SCLnt, probably as a result of increased nutrient gain as new recruits improve foraging behavior, diet selection, and digestive processing. Growth rates then decline to a size of about 90 cm SCLnt at which size green turtles approach maturity and growth rates slow as resource allocation is shifted from growth to reproduction. The slope of the decline becomes substantially steeper around 70 cm SCLnt, perhaps because of changes in habitat, diet selection, and food intake with increasing body size. A graph of predicted growth rates (cm/year) plotted against mean CL is presented in Fig. S2-2 (in Appendix S2). For discussion of size-specific growth dynamics, see Appendix S3.

Mean year of the growth increment also has a significant effect (p < .0001; Figure 2b). Growth rates increase from 1974 to 1999, although the 95% confidence interval is broad until ~1985. After 1999, growth rates exhibit a steady decline. This decline is not a result of changes in CL because, as noted above, covariates in the GAMM are conditioned on each other and mean CL values do not change over years. See Fig. S2-3 (in Appendix S2) for a graph of predicted growth rates (cm/year) plotted against mean year.

Growth rates differ significantly among diet categories (Figure 2c). Green turtles on seagrass diets grow more rapidly than green turtles on mixed seagrass/algae, algae, and omnivorous diets (p < .0001, p = .009, and p = .033, respectively). There are no significant differences among growth rates on seagrass/algae, algae, and omnivorous diets, although the difference between seagrass/algae and algae approaches significance (Figure 2c). For discussion of role of diet in growth dynamics, see Appendix S3.

The spatiotemporal interaction was significant for all epochs (p < .007; Fig. S2-1 in Appendix S2) and is confounded with all remaining heterogeneity in growth rates not accounted for by the six other covariates in our model (body size, mean year, duration, diet type, CL type, and project). Differences in growth rates indicated in Fig. S2-1 (in Appendix S2) by differences in color within an epoch and among epochs represent site-specific responses to other covariates not included in our model (e.g. food quality or quantity). If we could include other meaningful covariates in our model, the spatiotemporal plots presented here would probably present different patterns.

The nonsignificant fixed effects are duration (p = .076) and CL type (p > .235 for all comparisons). We set the range of recapture durations from 330 to 1,644 days so that duration would not affect the model (Figure 2d). Lack of significant difference among the four CL metrics (Figure 2e) justifies combining the growth data for the four measurement types.

Results of sea turtle growth studies are often presented for 10-cm carapace length size classes. To allow our results to be compared with other studies, we have provided these values in Table S2-1 (in Appendix S2).

3.2 | Drivers of mean year effect

Annualized mean growth rates increase to a high value in 1999 and then decline by 26% to 2015 (Figure 3a). The correlation of this pattern with SST values from 1982 to 2015 is moderate (r = −.43 to −.54 with 0–11 year lags; Fig. S2-4 in Appendix S2). Correlation improves greatly when data are restricted to years with stable and declining growth starting in 1997 (r = −.94 for 0-year lag; Figure 4). There is an apparent threshold temperature between 25.9°C and 26.0°C below which growth rates tend to increase with increasing SST and above which growth rates decline as SST increases.

Annualized mean growth rates for all years (1974–2015) correlate strongly (r = .74) with annualized MEI with 2- to 4-year distributed lags (Figure 5). An inverse-precision weighted GAMM (Fig. S2-5 in Appendix S2) with 3-year lagged MEI accounts for ~52% of the variance in the annualized mean growth rates. We found a statistically significant 2-year lag between annualized MEI and annualized somatic growth rates using the Granger-causality test [VAR(p = 2) model was best fit for p ranging from 1:10, F-test = 93.1, df = c(1,66), p < .0001]. Forecasting performance declined rapidly with increasing lags 3–10. Thus, including MEI from 2 years prior significantly improves the forecast performance of predicting current somatic growth above and beyond just simply

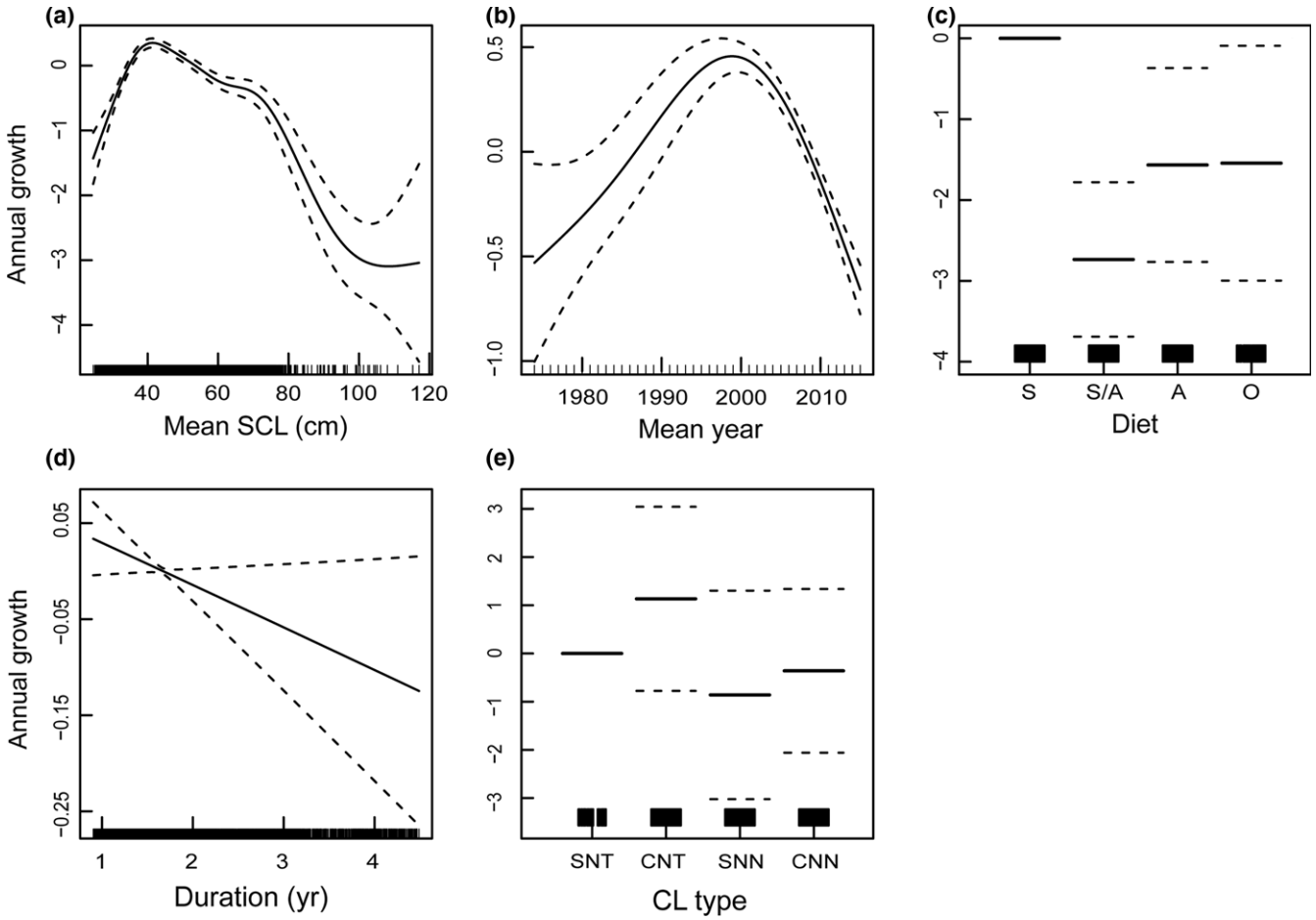

FIGURE 2 Graphical summary of GAMM analysis. The response variable (mean annual growth rate) is shown on the *y*-axis as a centered smoothed function scale to ensure valid pointwise 95% confidence bands and allow direct comparisons of effect strength among covariates. The covariate is shown on the *x*-axis: mean SCL (straight carapace length, cm) (a); mean year (b); diet (S is seagrass, S/A is seagrass and algae, A is algae, O is omnivorous) (c); duration (year) (d); CL (carapace length) measurement type (SNT is straight CL notch to tip, CNT is curved CL notch to tip, SNN is minimum straight CL, CNN is minimum curved CL, see Appendix S1) (e). Solid curves are the smoothing spline fits conditioned on all other covariates. Dashed lines are pointwise 95% confidence curves around the fits. All covariates are significant except duration and CL type. Rug plot indicates smaller sample sizes at large body size

using the growth rates themselves. This finding is consistent with the simpler lagged plot approach (Figure 5). Our results indicate that green turtle growth rates decrease with increasing SST above a threshold between 25.9°C and 26.0°C (Figures 3a,b and 4) and increase with increasing MEI (Figures 3a,c, and 5; Fig. S2-5 in Appendix S2).

4 | DISCUSSION

4.1 | Region-wide drivers of sea turtle growth declines

The significant regional decrease in green turtle growth rates after 1999 confirms that the pattern of decreasing growth rates in sea turtles beginning in the late 1990s and continuing to the present is consistent across trophic levels. Similar declines occur in annualized mean growth rates in two carnivorous species—West Atlantic hawksbills (Figure 3d) and North Atlantic loggerheads (Figure 3e,f)—following the highest growth rates in 1997. The growth functions for hawksbills (Figure 3d) and loggerheads (Figure 3e) were based on studies using capture–mark–recapture data and analyses similar to those in the present study (Bjorndal et al., 2013, 2016). The second loggerhead function (Figure 3f) was generated based on a very different approach using skeletochronology, different analyses, and a different loggerhead dataset (Avens et al., 2015) that reinforces the observed decline presented here. The different initial years of the declines among the three sea turtle species may represent different lag times in responding to environmental forces among the three species, but 1997 also falls within the 95% confidence interval for the highest growth rates in green turtles in 1999 (Figure 3a). One difference in these growth functions is the upturn in one of the loggerhead studies (Figure 3e) after 2007, but the confidence interval at that point would allow for a continued decline in growth rates.

Based on the similar growth dynamics among three sea turtle species across a trophic spectrum and on strong correlations with

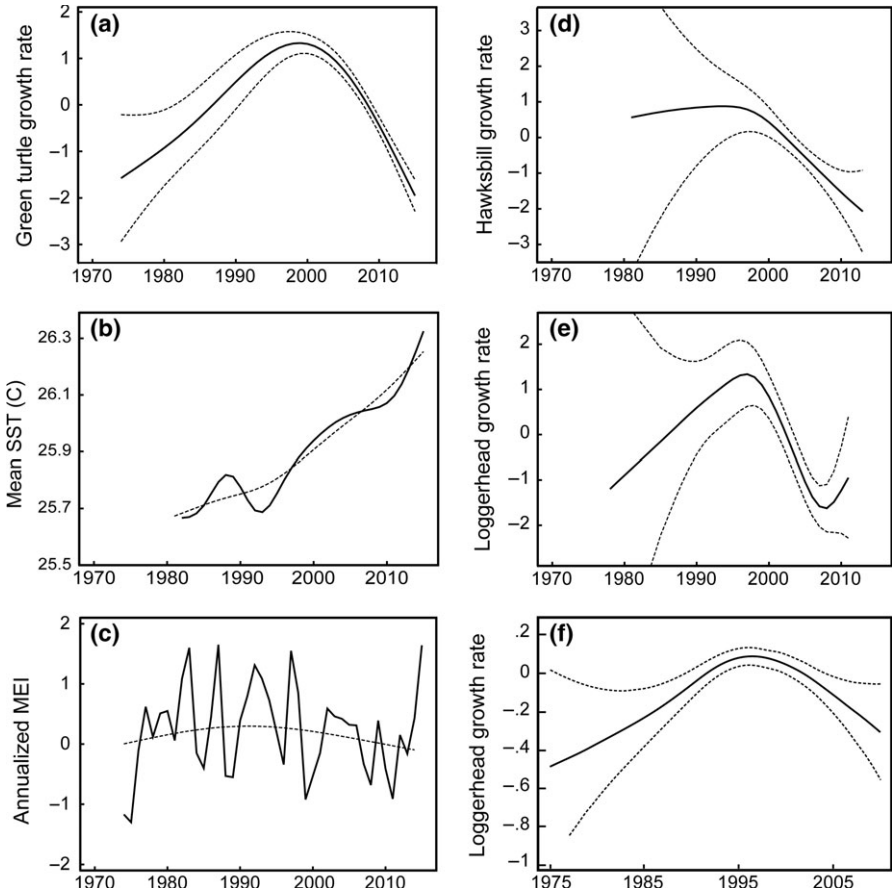

FIGURE 3 Annualized mean growth rates (standardized) for green turtles (a); annualized sea surface temperature (SST, °C) (b); annualized Multivariate El Niño Southern Oscillation Index (MEI) (c); annualized mean growth rates for hawksbills (standardized), modified from Bjorndal et al. (2016) (d); annualized mean growth rates (standardized) for loggerheads, modified from Bjorndal et al. (2013) (e); and loggerhead growth rates with centered smoothed GAMM function scale on the *y*-axis, modified from Avens et al. (2015) (f). For growth rates (a,d,e,f) solid lines are smoothing spline fits conditioned on all other covariates and dashed lines are pointwise 95% confidence curves around the fits. For SST and MEI (b,c) solid lines are annualized values and dashed lines are from GAMM analyses showing underlying annual trend; MEI data from 1950 to 1974 are not shown so that *x*-axes are consistent among graphs

MEI and SST, we conclude that the declining growth trajectories are most likely a result of the ERS that occurred in the late 1990s. The ERS is believed to be a result of the synergistic effect of two strong thermal processes: abrupt warming during the strong ENSO event of 1997/1998 and the intensification of warming rate over the last two to three decades (Beaugrand et al., 2015; IPCC, 2014; Martinson et al., 2008; Reid & Beaugrand, 2012; Wijffels et al., 2016). During this ERS, abrupt ecological changes occurred in the Atlantic from the North Sea to the Antarctic shelf, including substantial loss of Antarctic sea ice, extreme global bleaching event of corals, and shifts in distribution and phenology in populations of phytoplankton, zooplankton, molluscs, echinoderms, fish, and seabirds (Beaugrand, McQuatters-Gollop, Edwards, & Goberville, 2013; Beaugrand et al., 2015; Hoegh-Guldberg et al., 2007; Luczak, Beaugrand, Jaffré, & Lenoir, 2011; Martinson et al., 2008; Ortega, Celentano, Finkl, & Defeo, 2013).

The correlation between MEI and the green turtle growth function is strong (r = .74) throughout the study period, whereas SST is moderately correlated (r = −.54) with the entire growth function but strongly negatively correlated (r = −.94) with the declining growth function in years following the El Niño year and above the threshold between 25.9 and 26.0°C. The cause of this threshold is not known. It does not appear to be a threshold for green turtle functioning (see discussion of thermal effects below) unless maximum SST values surpass the optimal thermal zone of green turtles in their habitats in years with an annualized value of 26°C.

The decline in hawksbill growth rates was also strongly correlated with warming SST in the Caribbean and declining MEI values, with a better fit with the latter (Bjorndal et al., 2016). The MEI and SST effects were attributed to indirect negative effects of rising temperatures on foraging habitats (primarily coral reefs) and prey organisms. Similar explorations of climatic indices were not conducted in the loggerhead growth study although water temperature was suggested as a primary driver for the decline in growth rates (Bjorndal et al., 2013).

4.2 | Multiple stressors

Effects of ERS can be reinforced and prolonged by synergistic interactions of multiple stressors (Conversi et al., 2015). The decline in

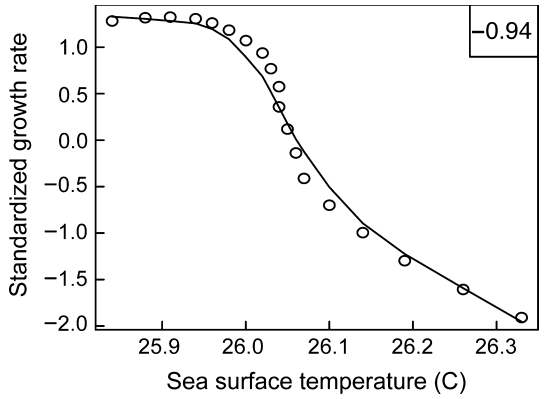

FIGURE 4 Annualized mean growth rates (standardized) of green turtles from 1997 to 2015 (open circles) against the annualized sea surface temperature (SST, °C) with no lag, solid line is the GAMM trend (see text). Correlation coefficient is in a box within the graph. Note the threshold between 25.9 and 26.0°C above which growth rates decline with increasing SST

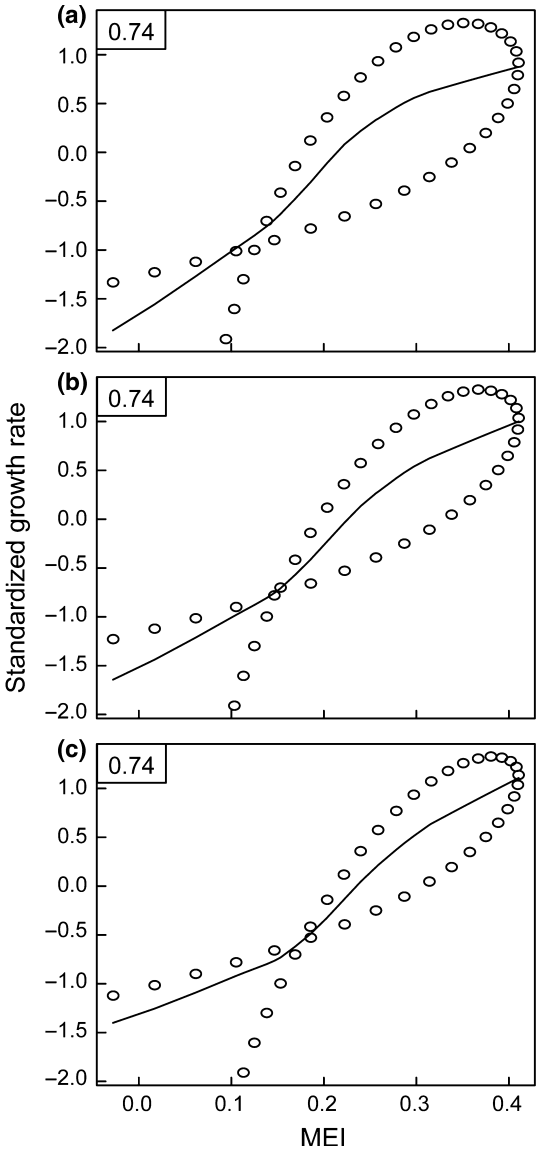

FIGURE 5 Annualized mean growth rates (standardized) of green turtles for 1974 to 2015 (open circles) lag-plotted against the annualized Multivariate El Niño Southern Oscillation Index (MEI) with 2-year lag (a), 3-year lag (b), and 4-year lag (c). Solid lines are the GAMM trends (see text). Correlation coefficients are in boxes within each graph

sea turtle growth rates may be a result of multiple stressors that are directly related to MEI or coincidental. Temperature can affect growth rates either directly, through physiological processes of sea turtles, or indirectly through effects on quality and quantity of food resources. Direct effects seem unlikely because the maximum SST values are well within the thermal activity range for sea turtles (Spotila, O'Connor, & Paladino, 1997). Therefore, any temperature influence would probably be indirect through effects on habitats and food resources, as reported for hawksbill growth rates (Bjorndal et al., 2016). Different aggregations of green turtles will not all exhibit the same temporal pattern in growth dynamics as the region-wide response in this study because of local differences in strength of stressors and the proximity of the green turtles to the edge of their thermal niche (Beaugrand et al., 2015).

In our study, 63% and 22% of growth increments are for turtles with primary diets of seagrasses (most commonly *Thalassia testudinum*) and seagrass/algae, respectively. Many reports exist of seagrasses living near their thermal maxima for both temperate and tropical species (Collier & Waycott, 2014; Pedersen, Colmer, Borum, Zavala-Perez, & Kendrick, 2016; Thomson et al., 2015). Increasing temperatures can have direct effects on physiological functions such as photosynthesis and reproduction (Bulthuis, 1987; Short & Neckles, 1999). Optimal temperatures for maximum productivity of *T. testudinum* range from 28°C to 31°C (Lee, Park, & Kim, 2007), and the threshold for *T. testudinum* under sustained exposure is ~33°C (Koch, Schopmeyer, Kyhn-Hansen, & Madden, 2007). Direct thermal effects on *T. testudinum* may seem unlikely with high values of monthly SST at 30°C in our study region. However, *T. testudinum* meadows often grow in shallow, protected waters that may experience water temperatures well above regional monthly SST and above the optimal thermal zone of the seagrass, especially at low tides (Collier & Waycott, 2014). Many indirect effects of increased temperatures on productivity, mortality, abundance, and distribution of seagrasses have been identified, including decrease in light penetration resulting from thermal-induced eutrophication, changes in salinity, and increased epiphytic algae, water depths, phytotoxins, and incidence of diseases (Koch et al., 2007; Short & Neckles, 1999).

Sea turtle foraging habitats are negatively impacted by many anthropogenic effects in addition to rising temperatures (Rees et al., 2016). The great increase in human populations in coastal areas (Norström et al., 2016) brings a plethora of threats to sea turtles and their habitats on continental shelves. Net human migration to coastal areas both globally and in areas of coral reefs remained constant in the 1970s and 1980s and increased greatly in the 1990s by factors of 2.7 and 5, respectively (Norström et al., 2016). The timing of this migration fits with the initiation of declines in sea turtle growth rates in the late 1990s and the dramatic decline in seagrass pastures.

Annual rates of loss of seagrass pastures have increased over the past decades, resulting in the loss of substantial seagrass area since the 1990s (Mcleod et al., 2011; Waycott et al., 2009). These are global seagrass losses, but within our study region seagrass loss has been substantial (Short & Wyllie-Echeverria, 1996). A network of 52 seagrass (primarily *T. testudinum*) sampling sites across the Greater Caribbean was monitored by CARICOMP from 1993 to the present (Van Tussenbroek et al., 2014). Of the 35 sites that allowed long-term monitoring, 15 (43%) had clear trends indicating environmental deterioration and 25 (71%) exhibited at least one of the six indicators of environmental deterioration (Van Tussenbroek et al., 2014).

Although some seagrass loss is from natural causes such as hurricanes, earthquakes, and foraging activities by a variety of species, the vast majority of loss is from anthropogenic activities. Industrial and agricultural run-off resulting in eutrophication, coastal infrastructure development, dredging, aquaculture development, algal blooms, trawling, and boat damage are some of the more important human activities that destroy seagrass pastures (Grech et al., 2012; Orth et al., 2006; Wells et al., 2015). The CARICOMP program identified increased terrestrial run-off of fertilizers, sewage, and sediments as the primary negative anthropogenic effects in the region (Linton & Fisher, 2004). The introduction of the invasive seagrass *Halophila stipulacea* in the eastern Caribbean is another potential stressor, and the combined environmental degradation may induce fibropapillomatosis, a green turtle disease that can reach high incidence (for discussion of both, see Appendix S3). As seagrass ecosystems decline, green turtles will shift to other diets of algae and invertebrates, if available. Based on our study, these diets support slower green turtle growth rates, and thus, the decline in growth rates will be exacerbated.

Anthropogenic degradation of foraging grounds of hawksbills and loggerheads are also well documented. Hawksbills are closely associated with coral reefs, and extent and health of reef habitats in the West Atlantic have suffered serious declines (references in Jackson, Donovan, Cramer, & Lam, 2014). Coral bleaching, acidification, and diseases interact synergistically with local stressors such as sedimentation, eutrophication, and overfishing to extend the effects of the ERS (Ateweberhan et al., 2013). Loggerheads are the most generalist of sea turtle species (Bolten, 2003) and occupy many habitats including seagrass pastures, hard bottom, and soft bottom habitats. Although diverse habitat use makes loggerheads less vulnerable to habitat destruction, they are not immune. Trawl fisheries and loggerhead foraging areas often overlap; trawl fishing drastically degrades bottom habitats and removes loggerhead prey (Bjorndal, 1997; National Research Council, 2002). All sea turtle habitats are also seriously degraded by accumulation of anthropogenic debris. Ingestion of marine debris by sea turtles has increased in the last few decades (Nelms et al., 2015) and can decrease nutrient gain in sea turtles through nutrient dilution which decreases growth rates (McCauley & Bjorndal, 1999).

Density dependence may be a factor in the decline of growth rates after the late 1990s because West Atlantic green turtle populations appear to be increasing in abundance (Chaloupka et al., 2008; García-Cruz et al., 2015; Weber et al., 2014). Also, as stated above, quality and quantity of foraging areas for sea turtles are declining; thus, lowering the population levels of green turtles at which density-dependent effects would be invoked. Evidence for density-dependent regulation of growth rates was reported for three green turtle study sites (The Bahamas, Florida, USA, and México; Bjorndal, Bolten, & Chaloupka, 2000; Kubis, Chaloupka, Ehrhart, & Bresette, 2009; Labrada-Martagón, Muñoz Tenería, Herrera-Pavón, & Negrete-Philippe, 2017), but no evidence of a density-dependent effect was found in a green turtle aggregation in Puerto Rico (Patrício, Diez, & van Dam, 2014). Density dependence cannot be the major driver because the three species of sea turtles would not simultaneously reach the population levels at which density dependence would begin to regulate somatic growth on a region-wide basis. Modern populations of hawksbills in the West Atlantic are a fraction of historical population sizes as a result of historic over-exploitation (McClenachan, Jackson, & Newman, 2006; Meylan & Donnelly, 1999). Increases in nest abundance for hawksbills reported for some areas in recent years have not been sufficient to recover these densities, even considering reductions in reef habitats (Campbell, 2014; NMFS & USFWS, 2013).

We conclude that the declining growth rates in sea turtles are most likely a result of an ERS that occurred in the late 1990s and exacerbated by the cumulative impacts of ongoing anthropogenic degradation of foraging habitats in the region. Determining the relative importance of individual stressors on growth rates is not possible at this time and deserves further research. Regardless of the mechanisms, the summary conclusion that productivity of sea turtles is lower at warmer temperatures is not good news in an age of warming seas.

ACKNOWLEDGEMENTS

All applicable institutional and/or national guidelines for care and use of animals were followed. Order of authorship, except the first four authors, was determined by number of growth increments contributed to this study. We thank colleagues from five sites who provided data that could not be included in our study because they fell below the cutoff: H. Barrios, N. Espinoza-Rodriguez, and N. Wildermann from the NGO Grupo de Trabajo en Tortugas Marinas del Golfo de Venezuela (Venezuela); S. Dunbar (Honduras); J. McNeill, L. Avens and A. Goodman Hall (North Carolina, USA); K. Holloway-Adkins (Florida, USA); and M. Tumlin (Louisiana, USA). Many of the data available for our study are the result of years of work by WIDECAST, the Wider Caribbean Sea Turtle Conservation Network and its Executive Director, K. Eckert. We also thank P. Eliazar for assistance with data management, J. Ogden, B. van Tussenbroek, and J. Cortés Núñez for making CARICOMP data available to us, and R. Johnson, A. Gulick and N. Constant for assistance with the seagrass literature review. We thank G. Hays and an anonymous reviewer for comments that improved the manuscript. Acknowledgments for individual projects are in Appendix S4. The authors have no conflicts of interest. Any use of trade, product, or firm names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

REFERENCES

Alheit, J. (2009). Consequences of regime shifts for marine food webs. *International Journal of Earth Sciences*, *98*, 261–268.

Alheit, J., & Bakun, A. (2010). Population synchronies within and between ocean basins: Apparent teleconnections and implications as to physical–biological linkage mechanisms. *Journal of Marine Systems*, *79*, 267–285.

Ateweberhan, M., Feary, D. A., Keshavmurthy, S., Chen, A., Schleyer, M. H., & Sheppard, C. R. C. (2013). Climate change impacts on coral reefs: Synergies with local effects, possibilities for acclimation, and management implications. *Marine Pollution Bulletin*, *74*, 526–539.

Avens, L., Goshe, L. R., Coggins, L., Snover, M. L., Pajuelo, M., Bjorndal, K. A., & Bolten, A. B. (2015). Age and size at maturation and adult stage duration for loggerhead sea turtles in the western North Atlantic. *Marine Biology*, *162*, 1749–1767.

Beaugrand, G., Conversi, A., Chiba, S., Edwards, M., Fonda-Umani, S., Greene, C., … Sugisaki, H. (2015). Synchronous marine pelagic regime shifts in the Northern Hemisphere. *Philosophical Transactions of the Royal Society B-Biological Sciences*, *370*, 20130272.

Beaugrand, G., McQuatters-Gollop, A., Edwards, M., & Goberville, E. (2013). Long-term responses of North Atlantic calcifying plankton to climate change. *Nature Climate Change*, *3*, 263–267.

Bjorndal, K. A. (1997). Foraging ecology and nutrition of sea turtles. In P. L. Lutz, & J. A. Musick (Eds.), *The biology of sea turtles* (pp. 199–231). Boca Raton, FL: CRC Press.

Bjorndal, K. A., & Bolten, A. B. (2008). Annual variation in source contributions to a mixed stock: Implications for quantifying connectivity. *Molecular Ecology*, *17*, 2185–2193.

Bjorndal, K. A., Bolten, A. B., & Chaloupka, M. Y. (2000). Green turtle somatic growth model: Evidence for density dependence. *Ecological Applications*, *10*, 269–282.

Bjorndal, K. A., Chaloupka, M., Saba, V. S., Diez, C. E., van Dam, R. P., Krueger, B. H., … Bolten, A. B. (2016). Somatic growth dynamics of West Atlantic hawksbill sea turtles: A spatio-temporal perspective. *Ecosphere*, *7*(5), e01279.

Bjorndal, K. A., Schroeder, B. A., Foley, A. M., Witherington, B. E., Bresette, M., Clark, D., … Bolten, A. B. (2013). Temporal, spatial, and body size effects on growth rates of loggerhead sea turtles (*Caretta caretta*) in the Northwest Atlantic. *Marine Biology*, *160*, 2711–2721.

Bolten, A. B. (2003). The loggerhead sea turtle: A most excellent fishe. In A. B. Bolten, & B. E. Witherington (Eds.), *Loggerhead sea turtles* (pp. 1–3). Washington, DC: Smithsonian Institution Press.

Bulthuis, D. A. (1987). Effects of temperature on photosynthesis and growth of seagrasses. *Aquatic Botany*, *27*, 27–40.

Campbell, C. L. (2014). *Conservation status of hawksbill turtles in the Wider Caribbean, Western Atlantic, and Eastern Pacific regions*. Arlington, VA: IAC Secretariat Pro Tempore. http://www.iacseaturtle.org/eng-docs/publicaciones/Conservacion-Carey-ing.pdf

Chaloupka, M., Bjorndal, K. A., Balazs, G. H., Bolten, A. B., Ehrhart, L. M., Limpus, C. J., … Yamaguchi, M. (2008). Encouraging outlook for recovery of a once severely exploited marine megaherbivore. *Global Ecology and Biogeography*, *17*, 297–304.

Chaloupka, M., & Limpus, C. J. (1997). Robust statistical modelling of hawksbill sea turtle growth rates (Southern Great Barrier Reef). *Marine Ecology Progress Series*, *146*, 1–8.

Collier, C. J., & Waycott, M. (2014). Temperature extremes reduce seagrass growth and induce mortality. *Marine Pollution Bulletin*, *83*, 483–490.

Conversi, A., Dakos, V., Gårdmark, A., Ling, S., Folke, C., Mumby, P. J., … Möllmann, C. (2015). A holistic view of marine regime shifts. *Philosophical Transactions of the Royal Society B-Biological Sciences*, *370*, 20130279.

Enders, W. (1995). *Applied econometric time series. Wiley series in probability and mathematical statistics*. New York, NY: John Wiley and Sons.

Fahrmeir, L., & Lang, S. (2001). Bayesian inference for generalised additive mixed models based on Markov random field priors. *Applied Statistics*, *50*, 201–220.

García-Cruz, M. A., Lampo, M., Peñaloza, C. L., Kendall, W. L., Solé, G., & Rodríguez-Clark, K. M. (2015). Population trends and survival of nesting green sea turtles *Chelonia mydas* on Aves Island, Venezuela. *Endangered Species Research*, *29*, 103–116.

Giannini, A., Chiang, J. C. H., Cane, M. A., Kushnir, Y., & Seager, R. (2001). The ENSO teleconnection to the tropical Atlantic Ocean: Contributions of the remote and local SSTs to rainfall variability in the tropical Americas. *Journal of Climate*, *14*, 4530–4544.

Gilman, E., Chaloupka, M., Peschon, J., & Ellgen, S. (2016). Risk factors for seabird bycatch in a pelagic longline tuna fishery. *PLoS ONE*, *11* (5), e0155477.

Gouirand, I., Moron, V., Hu, Z. Z., & Jha, B. (2014). Influence of the warm pool and cold tongue El Niños on the following Caribbean rainy season rainfall. *Climate Dynamics*, *42*, 919–929.

Grech, A., Chartrand-Miller, K., Erftemeijer, P., Fonseca, M., McKenzie, L., Rasheed, M., … Coles, R. (2012). A comparison of threats, vulnerabilities and management approaches in global seagrass bioregions. *Environmental Research Letters*, *7*, 024006.

Halpern, B. S., Frazier, M., Potapenko, J., Casey, K. S., Koenig, K., Longo, C., … Walbridge, S. (2015). Spatial and temporal changes in cumulative human impacts on the world's ocean. *Nature Communications*, *6*, 7615.

Ham, Y., Kug, J., Park, J., & Jin, F. (2013). Sea surface temperature in the north tropical Atlantic as a trigger for El Niño/Southern Oscillation events. *Nature Geoscience*, *6*, 112–116.

Hoegh-Guldberg, O., Mumby, P. J., Hooten, A. J., Steneck, R. S., Greenfield, P., Gomez, E., … Hatziolos, M. E. (2007). Coral reefs under rapid climate change and ocean acidification. *Science*, *318*, 1737–1742.

Hopkins-Murphy, S. R., Owens, D. W., & Murphy, T. M. (2003). Ecology of immature loggerheads on foraging grounds and adults in internesting habitat in the eastern United States. In A. B. Bolten, & B. E. Witherington (Eds.), *Loggerhead sea turtles* (pp. 79–92). Washington, DC: Smithsonian Institution Press.

IPCC (2014). *Climate change 2014: Synthesis report, summary for policy makers*. Geneva, Switzerland: IPCC.

Jackson, J. B. C., Donovan, M. K., Cramer, K. L., & Lam, V. V., editors (2014). *Status and trends of Caribbean coral reefs: 1970–2012*. Gland, Switzerland: Global Coral Reef Monitoring Network, IUCN.

Jackson, J. B. C., Kirby, M. X., Berger, W. H., Bjorndal, K. A., Botsford, L. W., Bourque, B. J., … Warner, R. R. (2001). Historical overfishing and the recent collapse of coastal ecosystems. *Science*, *293*, 629–638.

Kammann, E., & Wand, M. (2003). Geoadditive models. *Applied Statistics*, *52*, 1–18.

Koch, M. S., Schopmeyer, S., Kyhn-Hansen, C., & Madden, C. J. (2007). Synergistic effects of high temperature and sulfide on tropical seagrass. *Journal of Experimental Marine Biology and Ecology*, *341*, 91–101.

Krueger, B., Chaloupka, M., Leighton, P., Dunn, J., & Horrocks, J. (2011). Somatic growth rates for a hawksbill turtle population in coral reef habitat around Barbados. *Marine Ecology Progress Series*, *432*, 269–276.

Kubis, S., Chaloupka, M., Ehrhart, L., & Bresette, M. (2009). Growth rates of juvenile green turtles *Chelonia mydas* from three ecologically distinct foraging habitats along the east central coast of Florida, USA. *Marine Ecology Progress Series*, *389*, 257–269.

Labrada-Martagón, V., Muñoz Tenería, F. A., Herrera-Pavón, R., & Negrete-Philippe, A. (2017). Somatic growth rates of immature green turtles *Chelonia mydas* inhabiting the foraging ground Akumal Bay in the Mexican Caribbean Sea. *Journal of Experimental Marine Biology and Ecology*, *487*, 68–78.

Lee, K. S., Park, S. R., & Kim, Y. K. (2007). Effects of irradiance, temperature, and nutrients on growth dynamics of seagrasses: A review. *Journal of Experimental Marine Biology and Ecology*, *350*, 144–175.

Linton, D., & Fisher, T., editors (2004). CARICOMP – Caribbean Coastal Marine Productivity Program: 1993-2003. Caribbean Coastal Marine Productivity Program, Kingston, Jamaica.

Lotze, H. K., Lenihan, H. S., Bourque, B. J., Bradbury, R. H., Cooke, R. G., Kay, M. C., … Jackson, J. B. C. (2006). Depletion, degradation, and recovery potential of estuaries and coastal seas. *Science*, *312*, 1806–1809.

Luczak, C., Beaugrand, G., Jaffré, M., & Lenoir, S. (2011). Climate change impact on Balearic shearwater through a trophic cascade. *Biology Letters*, *7*, 702–705.

Mariusz, M. (2015). A review of the Granger-causality fallacy. *The Journal of Philosophical Economics*, *8*, 86–105.

Marra, G., Miller, D., & Zanin, L. (2012). Modelling the spatiotemporal distribution of the incidence of resident foreign population. *Statistica Neerlandica*, *66*, 133–160.

Martinson, D. G., Stammerjohn, S. E., Iannuzzi, R. A., Smith, R. C., & Vernet, M. (2008). Western Antarctic Peninsula physical oceanography and spatio–temporal variability. *Deep-Sea Research II*, *55*, 1964–1987.

Mazzarella, A., Giuliacci, A., & Scafetta, N. (2013). Quantifying the Multivariate ENSO Index (MEI) coupling to CO_2 concentration and to the length of day variations. *Theoretical Applied Climatology*, *111*, 601–607.

McCauley, S. J., & Bjorndal, K. A. (1999). Conservation implications of dietary dilution from debris ingestion: Sublethal effects in post-hatchling loggerhead sea turtles. *Conservation Biology*, *13*, 925–929.

McCauley, D. J., Pinsky, M. L., Palumbi, S. R., Estes, J. A., Joyce, F. H., & Warner, R. R. (2015). Marine defaunation: Animal loss in the global ocean. *Science*, *347*, 1255641.

McClenachan, L., Jackson, J. B. C., & Newman, M. J. H. (2006). Conservation implications of historic sea turtle nesting beach loss. *Frontiers in Ecology and the Environment*, *4*, 290–296.

Mcleod, E., Chmura, G. L., Bouillon, S., Salm, R., Björk, M., Duarte, C. M., … Silliman, B. R. (2011). A blueprint for blue carbon: Toward an improved understanding of the role of vegetated coastal habitats in sequestering CO_2. *Frontiers in Ecology and the Environment*, *9*, 552–560.

Meylan, A. B., & Donnelly, M. (1999). Status justification for listing the hawksbill turtle (*Eretmochelys imbricata*) as critically endangered on the 1996 IUCN Red List of Threatened Animals. *Chelonian Conservation and Biology*, *3*, 200–224.

Musick, J. A., & Limpus, C. J. (1997). Habitat utilization and migration in juvenile sea turtles. In P. L. Lutz, & J. A. Musick (Eds.), *The biology of sea turtles* (pp. 137–164). Boca Raton, FL: CRC Press.

National Research Council (2002). *Effects of trawling and dredging on seafloor habitat*. Washington, DC: National Academies Press.

Nelms, S. E., Duncan, E. M., Broderick, A. C., Galloway, T. S., Godfrey, M. H., Hamann, M., … Godley, B. J. (2015). Plastic and marine turtles: A review and call for research. *ICES Journal of Marine Science*, *73*, 165–181.

NMFS & USFWS (2013). *Hawksbill sea turtle (Eretmochelys imbricata)* 5-year review: Summary and evaluation. Silver Spring, MD: National Marine Fisheries Service and U.S. Fish and Wildlife Service.

Norström, A. V., Nyström, M., Jouffray, J. B., Folke, C., Graham, N. A. J., Moberg, F., … Williams, G. J. (2016). Guiding coral reef futures in the Anthropocene. *Frontiers in Ecology and the Environment*, *14*, 490–498.

Ortega, L., Celentano, E., Finkl, C., & Defeo, O. (2013). Effects of climate variability on the morphodynamics of Uruguayan sandy beaches. *Journal of Coastal Research*, *29*, 747–755.

Orth, R. J., Carruthers, T. J. B., Dennison, W. C., Duarte, C. M., Fourqurean, J. W., Heck Jr, K. L., … Williams, S. L. (2006). A global crisis for seagrass ecosystems. *BioScience*, *56*, 987–996.

Patrício, R., Diez, C., & van Dam, R. P. (2014). Spatial and temporal variability of immature green turtle abundance and somatic growth in Puerto Rico. *Endangered Species Research*, *23*, 51–62.

Pedersen, O., Colmer, T. D., Borum, J., Zavala-Perez, A., & Kendrick, G. A. (2016). Heat stress of two tropical seagrass species during low tides – Impact on underwater net photosynthesis, dark respiration and diel in situ internal aeration. *New Phytologist*, *210*, 1207–1218.

Rees, A. F., Alfaro-Shigueto, J., Barata, P. C. R., Bjorndal, K. A., Bolten, A. B., Bourjea, J., … Godley, B. J. (2016). Are we working towards global research priorities for management and conservation of sea turtles? *Endangered Species Research*, *31*, 337–382.

Reid, P. C., & Beaugrand, G. (2012). Global synchrony of an accelerating rise in sea surface temperature. *Journal of the Marine Biological Association UK*, *92*, 1435–1450.

Rocha, J. C., Peterson, G. D., & Biggs, R. (2015). Regime shifts in the Anthropocene: Drivers, risks, and resilience. *PLoS ONE*, *10*(8), e0134639.

Short, F. T., & Neckles, H. A. (1999). The effects of global climate change on seagrasses. *Aquatic Botany*, *63*, 169–196.

Short, F. T., & Wyllie-Echeverria, S. (1996). Natural and human induced disturbance of seagrasses. *Environmental Conservation*, *23*, 17–27.

Spillman, C. M., Alves, O., & Hudson, D. A. (2011). Seasonal prediction of thermal stress accumulation for coral bleaching in the tropical oceans. *Monthly Weather Review*, *139*, 317–331.

Spotila, J. R., O'Connor, M. P., & Paladino, F. V. (1997). Thermal biology. In P. L. Lutz, & J. A. Musick (Eds.), *The biology of sea turtles* (pp. 297–314). Boca Raton, FL: CRC Press.

Stoffer, D. (2014). astsa: Applied statistical time series analysis. R package version 1.3. http://CRAN.R-project.org/package=astsa

Thomson, J. A., Burkholder, D. A., Heithaus, M. R., Fourqurean, J. W., Fraser, M. W., Statton, J., & Kendrick, G. A. (2015). Extreme temperatures, foundation species, and abrupt ecosystem change: An example from an iconic seagrass ecosystem. *Global Change Biology*, *21*, 1463–1474.

Triacca, U. (2005). Is Granger causality analysis appropriate to investigate the relationship between atmospheric concentration of carbon dioxide and global surface air temperature? *Theoretical and Applied Climatology*, *81*, 133–135.

van Tussenbroek, B. I., Cortés, J., Collin, R., Fonseca, A. C., Gayle, P. M. H., Guzmán, H. M., … Weil, E. (2014). Caribbean-wide, long-term study of seagrass beds reveals local variations, shifts in community structure and occasional collapse. *PLoS ONE*, *9*(3), e90600.

Wallace, B. P., DiMatteo, A. D., Hurley, B. J., Finkbeiner, E. M., Bolten, A. B., Chaloupka, M. Y., … Mast, R. B. (2010). Regional management units for marine turtles: A novel framework for prioritizing conservation and research across multiple scales. *PLoS ONE*, *5*(12), e15465.

Waycott, M., Duarte, C. M., Carruthers, T. J. B., Orth, R. J., Dennison, W. C., Olyarnik, S., … Williams, S. L. (2009). Accelerating loss of seagrasses across the globe threatens coastal ecosystems. *Proceedings of the National Academy of Sciences of the United States of America*, *106*, 12377–12381.

Weber, S. B., Weber, N., Ellick, J., Avery, A., Frauenstein, R., Godley, B. J., … Broderick, A. C. (2014). Recovery of the South Atlantic's largest green turtle nesting population. *Biodiversity Conservation*, *23*, 3005–3018.

Wells, M. L., Trainer, V. L., Smayda, T. J., Karlson, B. S. O., Trick, C. G., Kudela, R. M., … Cochlan, W. P. (2015). Harmful algal blooms and climate change: Learning from the past and present to forecast the future. *Harmful Algae*, *49*, 68–93.

Wijffels, S., Roemmich, D., Monselesan, D., Church, J., & Gilson, J. (2016). Ocean temperatures chronicle the ongoing warming of earth. *Nature Climate Change*, *6*, 116–118.

Wood, S. (2006). *Generalized additive models: An introduction with R*. Boca Raton, FL: Chapman and Hall/CRC.

Wood, S., & Scheipl, F. (2014). gamm4: Generalized additive mixed models using mgcv and lme4. R package version 0.2-3. http://CRAN.R-project.org/package=gamm4

Young, J. W., Hunt, B. P. V., Cook, T. R., Llopiz, J. K., Hazen, E. L., Pethybridge, H. R., ... Choy, C. A. (2015). The trophodynamics of marine top predators: Current knowledge, recent advances and challenges. *Deep-Sea Research II*, *113*, 170–187.

SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

How to cite this article: Bjorndal KA, Bolten AB, Chaloupka M, et al. Ecological regime shift drives declining growth rates of sea turtles throughout the West Atlantic. *Glob Change Biol*. 2017;00:1–13. https://doi.org/10.1111/gcb.13712

Supporting Information

Ecological regime shift drives declining growth rates of sea turtles throughout the West Atlantic

Karen A. Bjorndal, Alan B. Bolten, Milani Chaloupka, Vincent S. Saba, Cláudio Bellini, Maria A. G. Marcovaldi, Armando J. B. Santos, Luis Felipe Wurdig Bortolon, Anne B. Meylan, Peter A. Meylan, Jennifer Gray, Robert Hardy, Beth Brost, Michael Bresette, Jonathan C. Gorham, Stephen Connett, Barbara Van Sciver Crouchley, Mike Dawson, Deborah Hayes, Carlos E. Diez, Robert P. van Dam, Sue Willis, Mabel Nava, Kristen M. Hart, Michael S. Cherkiss, Andrew G. Crowder, Clayton Pollock, Zandy Hillis-Starr, Fernando A. Muñoz Tenería, Roberto Herrera-Pavón, Vanessa Labrada-Martagón, Armando Lorences, Ana Negrete-Philippe, Margaret M. Lamont, Allen M. Foley, Rhonda Bailey, Raymond R. Carthy, Russell Scarpino, Erin McMichael, Jane A. Provancha, Annabelle Brooks, Adriana Jardim, Milagros López-Mendilaharsu, Daniel González-Paredes, Andrés Estrades, Alejandro Fallabrino, Gustavo Martínez-Souza, Gabriela M. Vélez-Rubio, Ralf H. Boulon, Jr., Jaime A. Collazo, Robert Wershoven, Vicente Guzmán Hernández, Thomas B. Stringell, Amdeep Sanghera, Peter B. Richardson, Annette C. Broderick, Quinton Phillips, Marta Calosso, John A. B. Claydon, Tasha L. Metz, Amanda L. Gordon, Andre M. Landry, Jr., Donna J. Shaver, Janice Blumenthal, Lucy Collyer, Brendan J. Godley, Andrew McGowan, Matthew J. Witt, Cathi L. Campbell, Cynthia J. Lagueux, Thomas L. Bethel and Lory Kenyon

Appendix S1. Supplemental Methods

Treatment of various carapace length measurements

The most common measure of body size in sea turtles is carapace length (CL). In our study, four different CL metrics were used in the 30 projects: straight carapace length notch to tip (SCLnt), straight carapace length minimum (SCLmin), curved carapace length notch to tip (CCLnt), and curved carapace length minimum (CCLmin) (Bolten, 1999). For those projects that measured more than one CL metric for each turtle, authors provided all measures of CL recorded for individual turtles to allow for conversions among different metrics. CL was measured with calipers for two straight CL (SCL) metrics: SCLnt and SCLmin (Fig. S1-1). CL was measured with flexible measuring tapes for two curved CL (CCL) metrics: CCLnt and CCLmin (Fig. S1-1).

In our GAMM analyses, we calculated growth increments based on any of the four metrics. In all cases, the same CL metric was used for initial and recapture CL. Each of the growth increments was coded by the type of CL metric (= CL type), and CL type was included as a factor in the model to test for effects of using different metrics.

To include body size in the GAMM analyses, a consistent mean CL metric is needed for all growth increments. We use SCLnt because it is the most common metric (n = 3799 growth increments), compared with SCLmin (n = 911), CCLnt (n = 1450), and CCLmin (n = 41). For turtles without SCLnt measurements, we use conversion equations based on data in our study to

estimate SCLnt from each of the other three CL measurements (Table S1-1). We use those equations to generate SCLnt values for each capture, and calculate the mean CL value for GAMM analyses. We also use the SCLnt values to calculate mean growth rates for the 10-cm SCLnt size classes in Table S2-1 in Appendix S2. Linear regressions were conducted in S-Plus software (TIBCO Spotfire S+ Version 8.2.0).

Fig. S1-1. The anterior and posterior pairs of anatomical points for four carapace length measurements. (A) Minimum straight carapace length (SCLmin) and minimum curved carapace length (CCLmin) are measured from the anterior point at midline (nuchal scute) to the posterior notch at midline between the supracaudals. (B) Straight carapace length notch to tip (SCLnt) and curved carapace length notch to tip (CCLnt) are measured from the anterior point at midline (nuchal scute) to the posterior tip of the supracaudals. Used with permission from Bolten (1999).

Table S1-1. Conversions among carapace length measurements.

Equation	Adjusted R^2	*n*	*P*
SCLnt = 1.0138*SCLmin + 0.238	0.999	4270	< 0.0001
SCLnt = 0.9698*CCLnt – 0.5385	0.989	1720	< 0.0001
SCLnt = 0.9572*CCLmin + 0.361	0.996	3475	< 0.0001

Notes: *n* is sample size; SCLnt is straight carapace length notch to tip, SCLmin is minimum SCL, CCLnt is curved carapace length notch to tip, CCLmin is minimum CCL.

Additional Granger-causality methods

If forecasting performance is improved, then MEI would be considered to Granger-cause growth rates. It was not possible to include MEI as an informative covariate directly in the geoadditive GAMM models because the growth rate metric and MEI are measured on completely different

time scales. So it must be assessed as a product of the GAMM-modelled output. Therefore we explored Granger-causality between the expected GAMM-modelled annualized growth rates and annualized MEI using a vector autoregressive modelling approach with variable lags (p) known as a VAR-p model (Enders, 1995). We implemented this approach using the *vars* package for R (Pfaff, 2008) with a robust variance-covariance matrix estimator (Zeileis, 2006) for the Granger test of any dynamic linear relationship between MEI and somatic growth. Stationarity for both time series was ensured using second differencing determined from KPSS unit root tests (Kwiatkowski *et al.*, 1992), although the results are similar irrespective of using a wide range of differencing including no differencing. The KPSS unit root tests were implemented using the *forecast* package for R (Hyndman & Khandakar, 2008). The lag order for the VAR-p with constant deterministic regressor (Enders, 1995) was assessed by estimating a series of models with lag order p ranging from 1 to 10 years – again the results were found to be independent of the deterministic regressor used such as trend, constant or both (Enders, 1995).

References for Appendix S1

Bolten AB (1999) Techniques for measuring sea turtles. In: *Management and Research Techniques for the Conservation of Sea Turtles* (eds. Eckert KL, Bjorndal KA, Abreu Grobois FA, Donnelly M), pp. 110‑114, IUCN, Gland, Switzerland.

Enders W (1995) Applied Econometric Time Series. Wiley Series in Probability and Mathematical Statistics. John Wiley and Sons, New York.

Hyndman R, Khandakar Y (2008) Automatic time series forecasting: the forecast package for R. Journal of Statistical Software, **26**, 1-22.

Kwiatkowski D, Phillips P, Schmidt P, Shin Y (1992) Testing the null hypothesis of stationarity against the alternative of a unit root. Journal of Econometrics, **54**, 159-178.

Pfaff B (2008) VAR, SVAR and SVEC models: implementation within R package vars. Journal of Statistical Software, **27**, 1-32.

Zeileis A (2006) Object-oriented computation of sandwich estimators. Journal of Statistical Software, **16**, 1–16.

Appendix S2. Supplemental Results

Fig. S2-1. Spatio-temporal interaction plots for the four epochs. All are significant (P < 0.007) and account for all remaining heterogeneity in growth rates not attributed to the six other covariates in our model (mean CL, mean year, duration, diet type, CL type, and project). The color scale is the centered GAM scale as for all the GAMM plots (e.g., Fig. 2) to allow comparison among plots. Red = lowest growth rates; beige = highest growth rates.

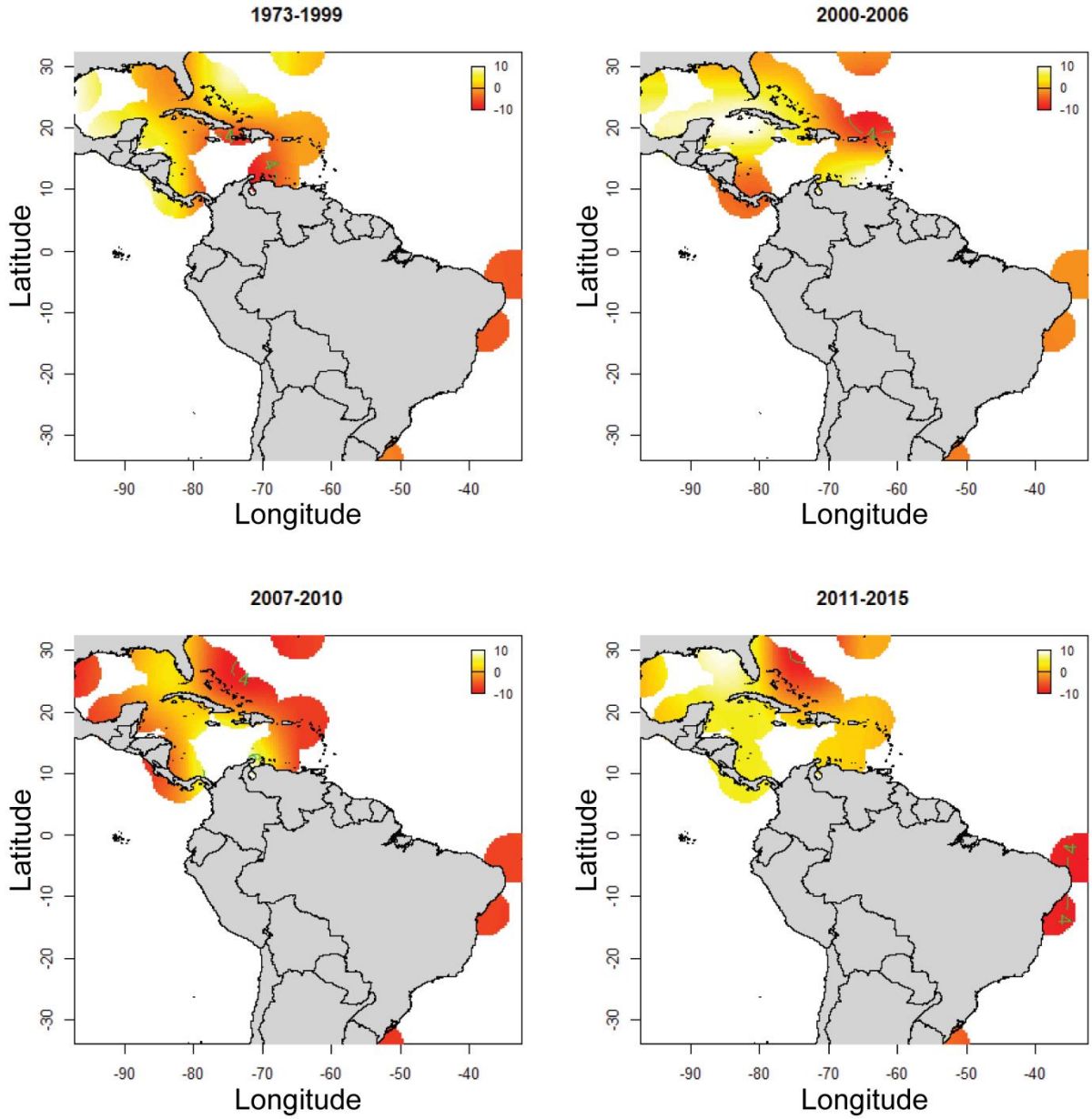

Fig. S2-2. Straight carapace length (SCL) growth rate (cm/yr) predicted by the GAMM analysis plotted against the mean SCL (cm) of each growth increment. Number of growth increments = 6201.

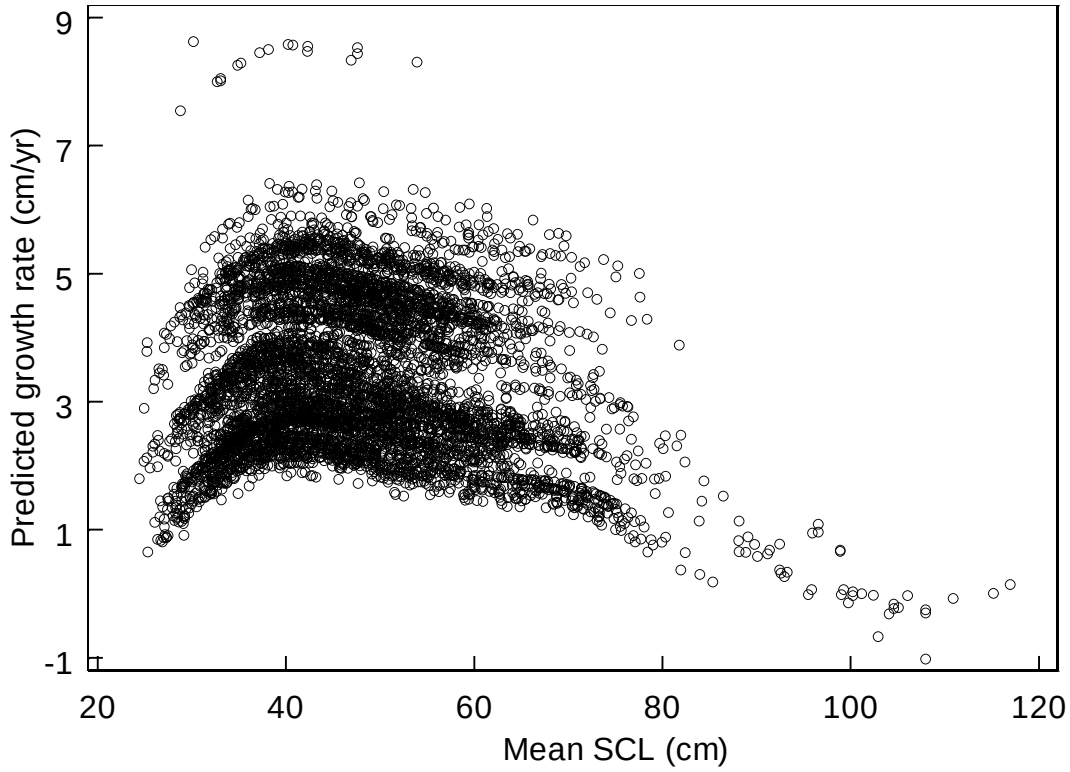

Fig. S2-3. Straight carapace length growth rate (cm/yr) predicted by the GAMM analysis plotted against the mean year of each growth increment. Number of growth increments = 6201.

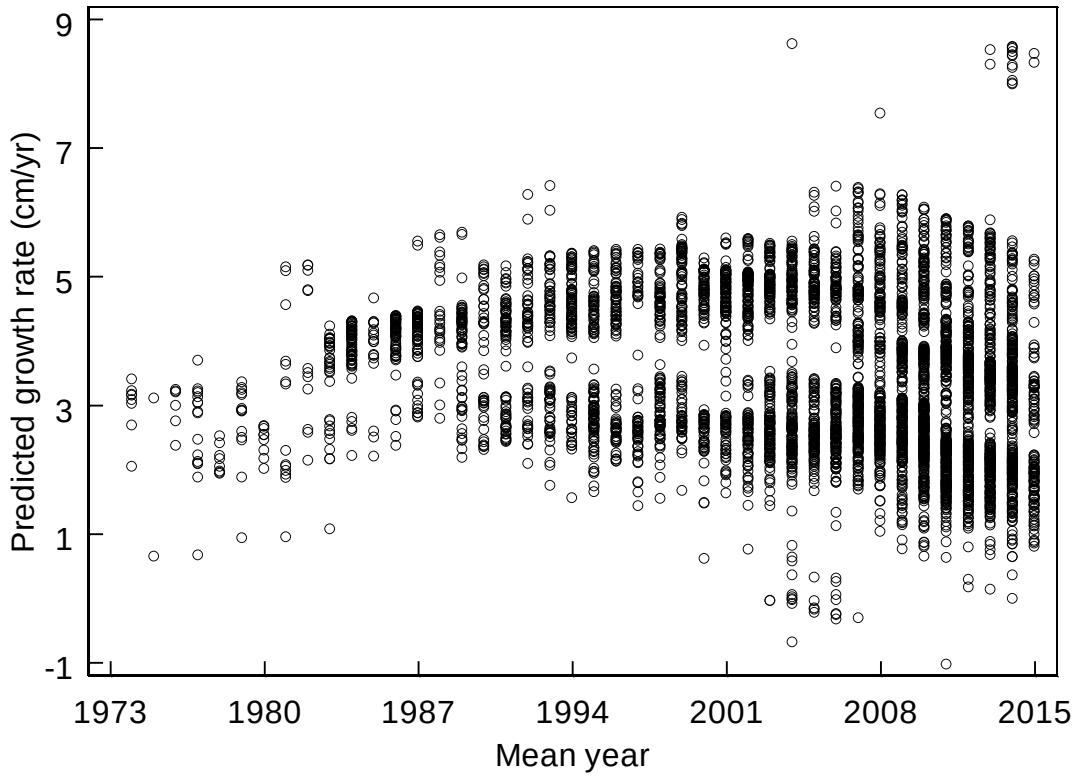

Fig. S2-4. Annualized mean growth rates and sea surface temperature. GAMM trends (solid lines) of annualized mean growth rates (standardized) of green turtles (open circles) lag-plotted against the annualized sea surface temperature (SST, °C) from 1982 to 2015 with 0- to 11-yr lags. Correlation coefficients are in boxes within each graph.

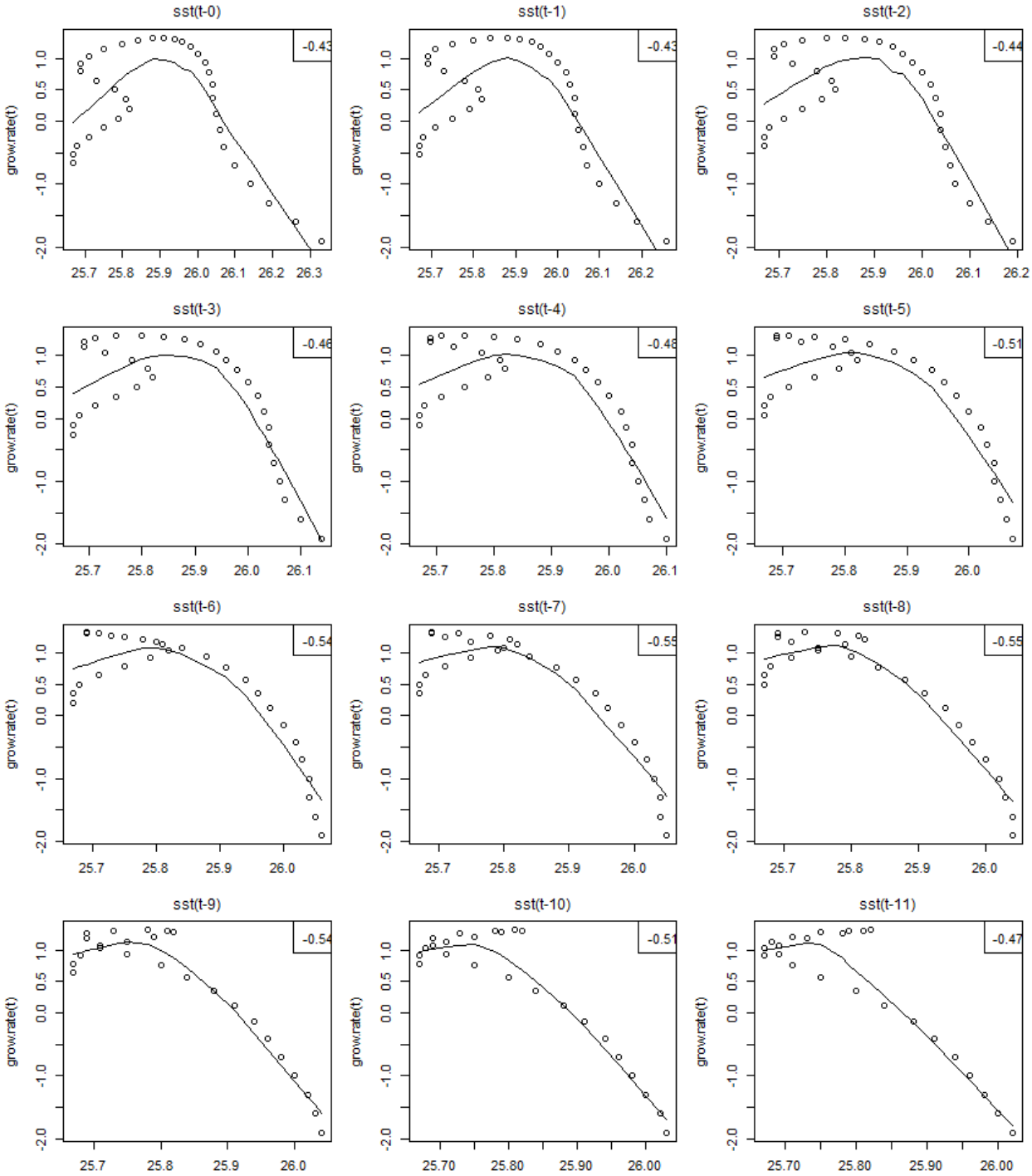

Fig. S2-5. Predicted year-specific standardized straight carapace length growth rates as a function of 3-yr lagged annualized Multivariate El Niño Southern Oscillation Index (MEI). The precision of each estimate is proportional to dot-size. The shaded area is bounded by 95% pointwise confidence curves.

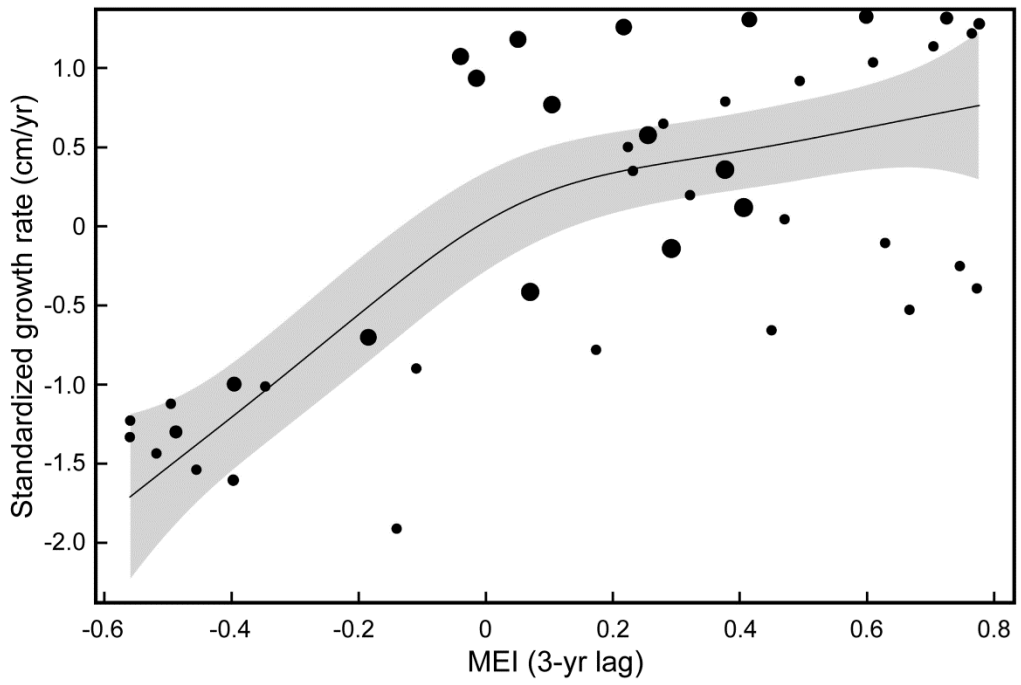

Table S2-1. Growth rates for 10-cm size classes

Results of sea turtle growth studies are often presented for 10-cm carapace length size classes. To allow our results to be compared with other studies, we have provided the values in Table S2-1. These growth rates should be interpreted with caution because they have not been conditioned on the other significant covariates (mean year, diet, spatio-temporal interaction) in our model. Mean year, diet, spatio-temporal distribution of growth increments may not be equally represented among the 10-cm size classes.

Table S2-1. Growth rates (cm/yr) for 10-cm straight carapace length notch to tip (SCLnt) size classes based on the dataset with 330 to 1644 d durations (n = 6201). n is sample size; SD is standard deviation.

Size class (cm)	*n*	Mean	Range	SD
20.0 – 29.9	152	2.3	-0.2 to 7.7	1.8
30.0 – 39.9	1552	3.4	-0.9 to 11.6	2.1
40.0 – 49.9	2105	3.8	-0.7 to 11.9	2.1
50.0 – 59.9	1487	3.5	-0.9 to 11.9	1.8
60.0 – 69.9	647	2.9	-0.2 to 9.2	1.5
70.0 – 79.9	199	2.3	-0.1 to 6.3	1.5
80.0 – 89.9	24	1.2	-0.2 to 3.3	1.0
90.0 – 99.9	19	0.2	-0.9 to 2.0	0.6
100.0 – 109.9	13	0	-0.7 to 0.6	0.3
110.0 – 119.9	3	-0.2	-0.9 to 0.4	0.7

Appendix S3. Supplemental Discussion

Size-specific growth dynamics

Body size (carapace length) has a very significant effect on growth rates in West Atlantic green turtles, which has been found in almost all studies of growth rates in sea turtles (Chaloupka & Limpus, 1997). However, the shape of the response function varies substantially among sea turtle populations (Chaloupka & Limpus, 1997; Bjorndal *et al.*, 2000). In West Atlantic green turtle aggregations, growth functions with increasing body size may be monotonic declining (e.g., Bjorndal *et al.*, 2000; Patrício *et al.*, 2014) or nonmonotonic with highest growth rates at relatively small sizes around 35 cm SCL (e.g., Kubis *et al.*, 2009) or at larger sizes around 50-55 cm SCL (e.g., Kubis *et al.*, 2009; Colman *et al.*, 2015).

Increasing growth rates over the smallest size range in our study from 25 to ~40 cm SCLnt (Fig. 2a) probably result from increased nutrient gain as new recruits improve foraging behavior, diet selection, and digestive processing. A shift from a diet of largely gelatinous prey in oceanic habitats (Bolten, 2003; Witherington *et al.*, 2012) to a largely herbivorous diet following recruitment to neritic habitats requires substantial changes in digestive processing and gut microbial populations (Bjorndal, 1997). An interval of limited nutrient gain while adapting to a new habitat and diet could be followed by a period of "catch-up" or compensatory growth during which turtles exhibit more rapid size-specific growth than if earlier nutrient gain had not been limited (Bjorndal *et al.*, 2003; Roark *et al.*, 2009).

Growth rates in green turtles larger than ~40 cm SCLnt in our study exhibit a consistent decline with two shifts in rate at about 70 cm and 90 cm SCLnt. Growth rates decline relatively slowly between 40 and 70 cm SCLnt and more rapidly between 70 and 90 cm SCLnt. Above 90 cm SCLnt growth greatly slows and approaches zero as green turtles approach and attain sexual maturity and shift allocation of nutrients from somatic growth to reproduction. The cause of the change in slope at 70 cm SCLnt is not known but may represent changes in habitat, diet selection, and food intake with increasing body size. Further studies on the foraging ecology of neritic green turtles are needed to understand more fully their size-specific growth dynamics.

Role of diet in growth dynamics

Green turtles feeding primarily on seagrasses (n = 3911 growth increments) have significantly higher growth rates than those feeding primarily on algae (n = 871), seagrass/algae mix (n = 1337), or an omnivorous diet (n = 82). Growth rates of green turtles feeding on the last three categories do not differ significantly. Within the seagrass category, the major species consumed is *Thalassia testudinum*, but other seagrass species, including *Halodule wrightii* and *Syringodium filiforme*, are also ingested (Guebert-Bartholo *et al.*, 2011; Stringell *et al.*, 2016).

Omnivorous diets might be expected to support more rapid growth because of the assumption that animal matter is more nutritious than plant matter to most animals. However, animal prey ingested by green turtles in neritic habitats, such as sponges, may provide nutrient gains equivalent to or lower than that of seagrasses. The sponge *Chondrilla caribensis* (= *nucula*) is one of the sponge species most commonly ingested by Atlantic green turtles

(Bjorndal, 1990; Stringell *et al.*, 2016). Green turtles digest organic matter and energy of *C. caribensis* to a significantly lesser extent than those of *T. testudinum*; apparent digestibility of nitrogen is similar between *C. caribensis* and *T. testudinum* (Bjorndal, 1990).

Variation in growth rates on different diets depends largely on differences in daily nutrient gain (daily intake of digestible nutrients or energy) on those diets. Nutrient gain in wild Atlantic green turtles feeding on natural diets has been quantified only for green turtles feeding on *T. testudinum* (Bjorndal *et al.*, 2000), as far as the authors are aware. Nutritional studies that quantify intake and digestibility on a range of natural diets so that daily gain of nutrients can be estimated are greatly needed to understand the role of sea turtles in the trophodynamics of marine systems.

Other stressors: invasive seagrass and fibropapillomatosis

Seagrass pastures and green turtle foraging may also be threatened by an invasive seagrass *Halophila stipulacea*, which is native to the Indo-Pacific, invaded the Mediterranean Sea in the late 1800s after the opening of the Suez Canal, was first identified in the West Atlantic in 2002 in Grenada, and has since spread rapidly in the eastern Caribbean (Willette *et al.*, 2014). *Halophila stipulacea* has probably not had a substantial role in declining growth rates of green turtles because of its relatively recent introduction and relatively restricted current range (parts of the eastern Caribbean). In addition, *H. stipulacea* is an important diet species for green turtles in the Indo-Pacific, and green turtles feed on it in the Caribbean (Becking *et al.*, 2014). Future studies should monitor effects of *H. stipulacea* on stability of seagrass beds and green turtle productivity.

Fibropapillomatosis (FP) is a widespread disease of green turtles that has attained incidence levels of 92% in some aggregations (Herbst, 1994). In the Atlantic, FP was first reported in green turtles from Florida in the late 1930s, but apparently only reached substantial levels in the early 1980s along the east coast of Florida (Herbst, 1994). After that time, FP reached epidemic levels in some areas in our study region (Patrício *et al.*, 2012). We did not include FP as a covariate in our growth models because monitoring for FP was too inconsistent throughout the spatial and temporal ranges of our study. Although it would seem likely that FP could have a substantial effect on growth rates, all studies to date have reported little or no effect. No effect of FP on growth rates of green turtles was found in two sites in Florida (Kubis *et al.*, 2009; Avens *et al.*, 2012), and at two sites in Puerto Rico (Patrício *et al.*, 2011, 2014). In Hawaii, growth rates were lower only in severe cases of the disease (Chaloupka & Balazs, 2005). In addition, body condition index did not differ significantly between green turtles with and without FP in Puerto Rico (Patrício *et al.*, 2016). Thus, we conclude that FP has at most a minor effect on growth rates in our study.

References for Appendix S3

Avens L, Goshe L, Harms C *et al.* (2012) Population characteristics, age structure, and growth dynamics of neritic juvenile green turtles in the northeastern Gulf of Mexico. Marine Ecology Progress Series, **458**, 213-229.

Becking LE, van Bussel TCJM, Debrot AO, Christianen MJA (2014) First record of a Caribbean green turtle (*Chelonia mydas*) grazing on invasive seagrass (*Halophila stipulacea*). Caribbean Journal of Science, **48**, 162-163.

Bjorndal KA (1990) Digestibility of the sponge *Chondrilla nucula* in the green turtle, *Chelonia mydas*. Bulletin of Marine Science, **47**, 567-570.

Bjorndal KA (1997) Foraging ecology and nutrition of sea turtles. In: *The Biology of Sea Turtles* (eds Lutz PL, Musick JA), pp. 199-231, CRC Press, Boca Raton, Florida.

Bjorndal KA, Bolten AB, Chaloupka MY (2000) Green turtle somatic growth model: evidence for density dependence. Ecological Applications, **10**, 269-282.

Bjorndal KA, Bolten AB, Dellinger T, Delgado C, Martins HR (2003) Compensatory growth in oceanic loggerhead sea turtles: response to a stochastic environment. Ecology, **84**, 1237-1249.

Bolten AB (2003) Variation in sea turtle life history patterns: neritic vs. oceanic developmental stages. In: *The Biology of Sea Turtles, volume II* (eds Lutz PL, Musick J, Wyneken J), pp. 243-257, CRC Press, Boca Raton, FL.

Chaloupka M, Balazs G (2005) Modelling the effect of fibropapilloma disease on the somatic growth dynamics of Hawaiian green sea turtles. Marine Biology, **147**, 1251‑1260.

Chaloupka M, Limpus CJ (1997) Robust statistical modelling of hawksbill sea turtle growth rates (Southern Great Barrier Reef). Marine Ecology Progress Series, **146**, 1‑8.

Colman MO, Patrício ARC, McGowan A, Santos AJB, Marcovaldi MÂ, Bellini C, Godley BJ (2015) Long‑term growth and survival dynamics of green turtles (*Chelonia mydas*) at an isolated tropical archipelago in Brazil. Marine Biology, **162**, 111-122.

Guebert-Bartholo FM, Barletta M, Costa MF, Monteiro-Filho ELA (2011) Using gut contents to assess foraging patterns of juvenile green turtles *Chelonia mydas* in the Paranaguá Estuary, Brazil. Endangered Species Research, **13**, 131–143.

Herbst LH (1994) Fibropapillomatosis of marine turtles. Annual Review of Fish Diseases, **4**, 389–425.

Kubis S, Chaloupka M, Ehrhart L, Bresette M (2009) Growth rates of juvenile green turtles *Chelonia mydas* from three ecologically distinct foraging habitats along the east central coast of Florida, USA. Marine Ecology Progress Series, **389**, 257−269.

Patrício AR, Velez-Zuazo X, Diez CE, Van Dam R, Sabat AM (2011) Survival probability of immature green turtles in two foraging grounds at Culebra, Puerto Rico. Marine Ecology Progress Series, **440**, 217−227.

Patrício AR, Herbst LH, Duarte A *et al.* (2012) Global phylogeography and evolution of the

chelonid fibropapilloma-associated herpesvirus. Journal of General Virology, **93**, 1035−1045.

Patrício R, Diez C, van Dam RP (2014) Spatial and temporal variability of immature green turtle abundance and somatic growth in Puerto Rico. Endangered Species Research, **23**, 51−62.

Patrício AR, Diez CE, van Dam RP, Godley BJ (2016). Novel insights into the dynamics of green turtle fibropapillomatosis. Marine Ecology Progress Series, **547**, 247–255.

Roark AM, Bjorndal KA, Bolten AB (2009) Compensatory responses to food restriction in juvenile green turtles (*Chelonia mydas*). Ecology, **90**, 2524–2534.

Stringell TB, Clerveaux WV, Godley BJ *et al.* (2016) Taxonomic distinctness in the diet of two sympatric marine turtle species. Marine Ecology, **37**, 1036–1049.

Willette DA, Chalifour J, Debrot AOD *et al.* (2014) Continued expansion of the trans-Atlantic invasive marine angiosperm *Halophila stipulacea* in the Eastern Caribbean. Aquatic Botany, **112**, 98– 102.

Witherington BE, Hirama S, Hardy R (2012) Young sea turtles of the pelagic Sargassum-dominated drift community: habitat use, population density, and threats. Marine Ecology Progress Series, **463**, 1-22.

Appendix S4. Acknowledgements for Individual Projects

For field assistance, logistics and funding, we thank:

Bahamas (Inagua): Samuel Nixon, James Nixon, Henry Nixon, Randolph Burrows, Lynn Gape, the Bahamas National Trust, Bahamas Department of Marine Resources, Morton Bahamas, Ltd, U.S. Coast Guard Airstation Clearwater, National Marine Fisheries Service, Disney Wildlife Conservation Fund

Bahamas (North and Central): Interns and students of the Cape Eleuthera Island School and Cape Eleuthera Institute, Meagan Gary, Rachel Miller, Edward Adderley, Ninive Espinoza, Melanie Esch, Luke Sundquist, Brittney Parker, Earthwatch Institute, Taylor Foundation, Cape Eleuthera Foundation

Bermuda: J. Burnett-Herkes, H. C. Frick, D. Godfrey, E. Harrison, J. Nisbett, C. O'Connor, G. Roth, I. D. Walker, R. Winchell, Bermuda Turtle Project annual course participants, local volunteers, Bermuda Aquarium Museum and Zoo staff, Bermuda Zoological Society staff and donors, The Atlantic Conservation Partnership, The Bermuda Government Department of Environment and Natural Resources, Chevron International Limited Bermuda, Eckerd College, Florida Fish & Wildlife Conservation Commission, the Helen Clay Frick Foundation, and the Sea Turtle Conservancy

Bonaire: WWF-NL (Wereld Nature Fonds – NL), EZ (Ministry of Economic Affairs NL), private and corporate funders http://www.bonaireturtles.org/wp/about-us/funding/, WIDECAST, Dive Friends Bonaire, Harbor Village Marina, STCB staff since 2003 and the ongoing support of many in-water volunteers over the years

Brazil (AdR, FdN, and PdF): All of the staff at Projeto TAMAR – Praia do Forte for their help with data collection and logistic support

México (Akumal & Campeche): Ana L. Talavera, Concepción Ortiz, Thierry M. Work, Alejandro Arenas Martinez, Julio Zurita Gutiérrez

Nicaragua: Data were collected under the auspices of the Nicaragua Sea Turtle Conservation Program of the Wildlife Conservation Society. Other assistance and support were provided by W.A. McCoy, Nicaragua fishers for their donations of live green turtles, Chelonia Institute, Chelonia Research Foundation – The Linnaeus Fund, DIPAL II Netherlands, Lerner-Gray Fund for Marine Research, National Marine Fisheries Service, Project Aware, and Tropical Conservation and Development Program University of Florida.

Panama: Inocencio (Chencho) Castillo, S. Duncan, T. Engstrom, D. Godfrey, C. Ordoñez. E. Possardt, A. Ruiz, R. Yeomans, the Wildlife Conservation Society, the Sea Turtle Conservancy, Busch Gardens Sea World Conservation Fund, the National Fish and Wildlife Foundation, the Explorers Club of New York, Andy and Molly Barnes, Eckerd

College, Florida Fish and Wildlife Conservation Commission, and the Smithsonian Tropical Research Institute

Puerto Rico (includes Culebra NWR): T. Tallevast, A. Tucker, J. Howell, R. DiRosa, A. Tucker, H. Morales, M. Oliveira, J. Glynn, K. Wolcott, and volunteers from Earthwatch

Turks and Caicos Islands: The Turks and Caicos Islands Turtle Project is a collaborative project between the Department of Environment and Coastal Resources (DECR), TCI; Marine Conservation Society (MCS), UK; University of Exeter, UK; Duke University, USA; and The School for Field Studies, Center for Marine Resource Studies (SFS-CMRS), TCI. It was established in November 2008 to assess marine turtle populations and their use in TCI with a view to improving the management of the Islands' turtle fishery. This work was funded by: Natural Environment Research Council CASE PhD studentship to T.B. Stringell, with the MCS as CASE partners (Ref: NE/F01385X/1). Simon and Anne Notley provided additional support and DECR and SFS-CMRS gave in-country support. We thank SFS-CMRS staff and students and Flora Kent for fieldwork assistance. Special thanks to the fishers of TCI who made the project possible.

USA, Florida (Dry Tortugas): USGS's Greater Everglades Priority Ecosystems Science (GEPES); permits for Dry Tortugas (PI Kristen M. Hart): National Marine Fisheries Service Endangered Species Permits 13307 and 17381, Marine Turtle Permit 176, and Dry Tortugas National Park permits DRTO-2016-SCI-0008, DRTO-2014-SCI-0004, DRTO-2012-SCI-0008, DRTO-2010-SCI-0009, DRTO-2008-SCI-0008

USA, Florida (East Coast projects): Betty Wershoven and Annalise Wershoven; Kennedy Space Center Ecological Program: D. Scheidt, R. Lowers, R. Cancro, M. Provancha, R. Bolt, E. Reyier, K. Holloway, C. Schumann, L. Phillips

USA, Florida (St Joseph Bay): Bruce Hagedorn at Eglin Air Force Base

USA, Texas (Laguna Madre): David Costa; F. Len Kenyon; Michael Coyne; Stacie Arms; Katie St. Clair; George Guillen; US Army Corps of Engineers; Texas Sea Grant Program; Carole Allen and Sea Turtle Restoration Project; Texas Parks and Wildlife Department; Texas State Aquarium Wildlife Care, Conservation and Research Fund; Sea Turtle, Inc.

USA, Texas (Mansfield Channel): Kristen Brennan, Darrell Echols, Paul Eubank and the many others that aided with field activities

US Virgin Islands: Brendalee Phillips, Phillipe Mayor, Kimberly Woody, Roy Pemberton Jr., and Jason Allen; US National Park Service Natural Resource Preservation Program (NRPP); permits (PI Kristen M. Hart): National Marine Fisheries Service Endangered Species Permit 16146 and Buck Island Reef National Monument permits BUIS-2012-SCI-0002, BUIS-2014-SCI-0009, BUIS-2011-SCI-0012